# CD MICROFLUIDICS FOR MOLECULAR DIAGNOSTICS

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# OUTLINE

- CDASA DIAGNOSTIC PLATFORM
  - CD MICROFLUIDICS FUNDAMENTALS
- CELLLYSIS

- DNA ARRAY HYBRIDIZATION
  - NEXT STEPS TOWARDS SAMPLE-TO-ANSWER SYSTEMS



SIEGRIST, ET AL, 2009

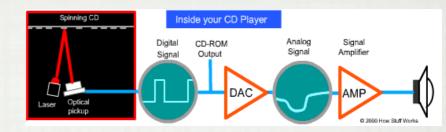


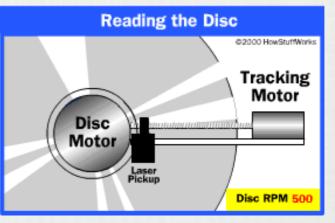


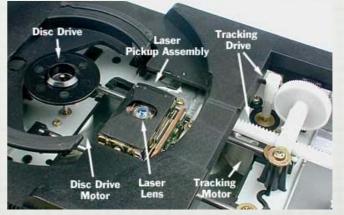
#### HOW DOES A CD PLAYER WORK?

THE FUNDAMENTAL JOB OF THE CD PLAYER IS TO FOCUS THE LASER ON A TRACK OF BUMPS. THE LASER BEAM PASSES THROUGH THE POLYCARBONATE CD MATERIAL AND REFLECTS OFF AN ALUMINUM LAYER AND HITS AN OPTO-ELECTRONIC DEVICE THAT DETECTS CHANGES IN LIGHT. THE BUMPS REFLECT LIGHT DIFFERENTLY THAN THE "LANDS" (THE REST OF THE ALUMINUM LAYER), AND THE OPTO-ELECTRONIC SENSOR DETECTS THAT CHANGE IN REFLECTIVITY.

A TRACKING MECHANISM MOVES THE LASER ASSEMBLY SO THAT THE LASER'S BEAM CAN FOLLOW THE SPIRAL TRACK. THE TRACKING SYSTEM HAS TO BE ABLE TO MOVE THE LASER AT MICRON RESOLUTIONS.





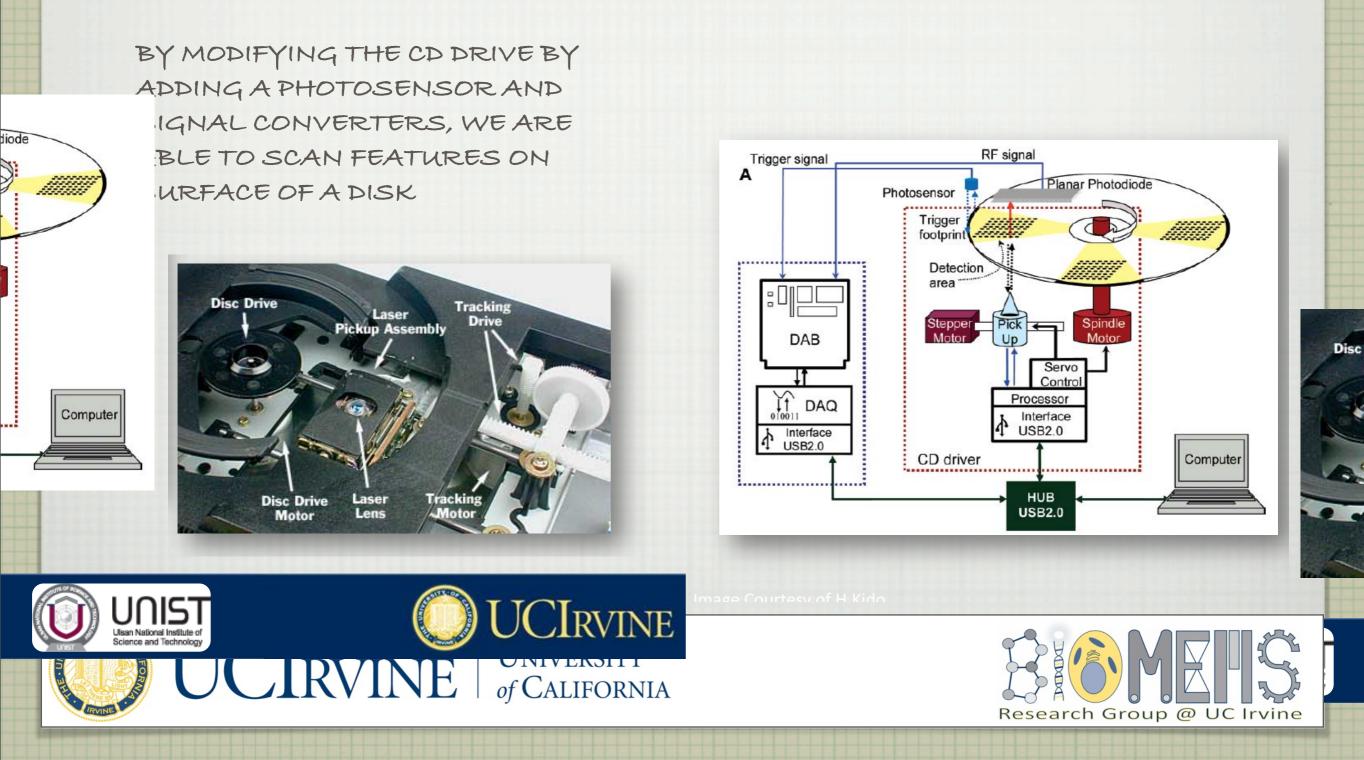


H. KIDO, J. ZOVAL





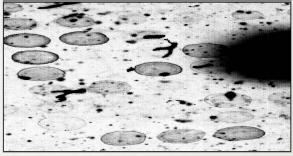
# CD AS A MICROSCOPE



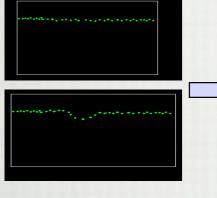
# CD AS A MICROSCOPE

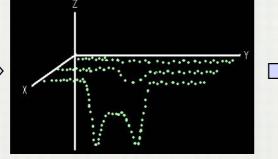
VOLTAGES READ BY PHOTO DETECTOR ARE RECONSTRUCTED INTO IMAGES WITH A/D CONVERTERS AND GRAPHICS SOFTWARE





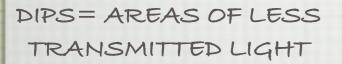
IMMUNDASSAYS

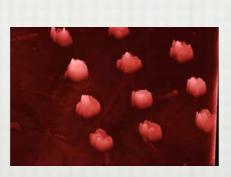












DNA ARRAYS



WHITE BLOOD CELLS





IMAGES COURTESY OF H.KIDO

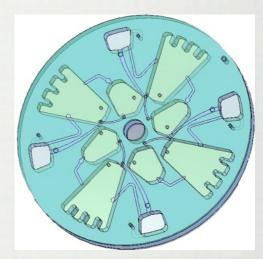




# CD AS A DIAGNOSTIC SYSTEM

#### ADVANTAGES OF CENTRIFUGAL MICROFLUIDICS

- DEPROGRAMMABLE CENTRIFUGE AUTOMATION OF SEVERAL PROCESSES ONTO SINGLE DISC
- COMPATIBLE WITH A WIDE RANGE OF SAMPLES
- NO FLUID CONTACT WITH PUMPING MECHANISM
- LIQUID AND VAPOR VALVING SOLUTIONS
- MULTIPLEXED DESIGNS FOR HIGH THROUGHPUT
- RAPID PROTOTYPING
- CHEAP, DISPOSABLE PLASTIC DEVICE
  - MASS PRODUCIBLE INJECTION MOLDING, HOT EMBOSSING

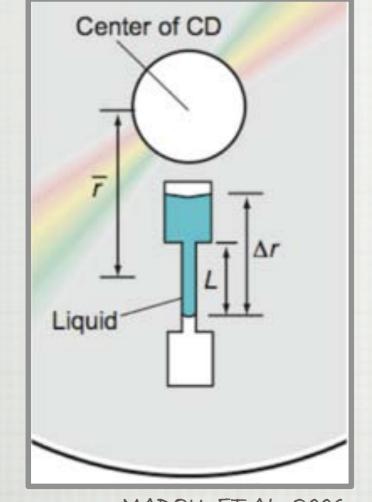








# CD MICROFLUIDICS FUNDAMENTALS



MADON, ET AL. 2006

CD FLUID PROPULSION IS ACHIEVED THROUGH CENTRIFUGALLY INDUCED PRESSURE AND DEPENDS ON ROTATION RATE, GEOMETRY AND LOCATION OF CHANNELS AND RESERVOIRS, AND FLUID PROPERTIES.

FLOW RATE, Q, IN A MICROFLUIDIC CD IS DESCRIBED BY:

 $Q = A D_h^2 \rho \omega^2 \overline{r} \Delta r / 32 \mu L$ 

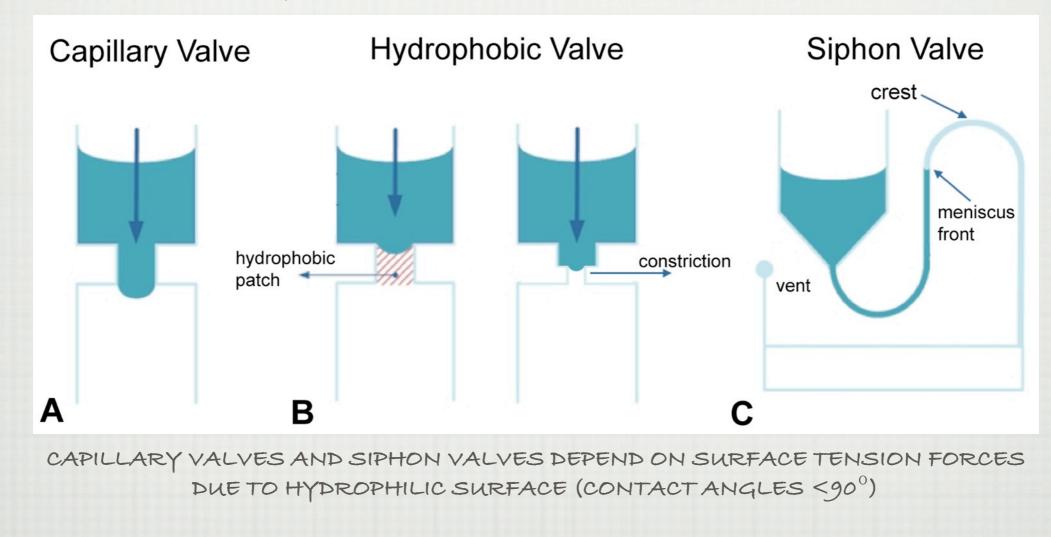
A is channel cross-sectional area
D<sub>h</sub> is hydraulic diameter
ρ is liquid density
ω is angular velocity of the spinning CD
r is the average distance of the liquid from the CD center
Δr is radial extent of the fluid
μ is fluid viscosity
L is the length of liquid in the channel

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# CD MICROFLUIDICS FUNDAMENTALS

LIQUID VALVING MECHANISMS FOR THE CD PLATFORM:

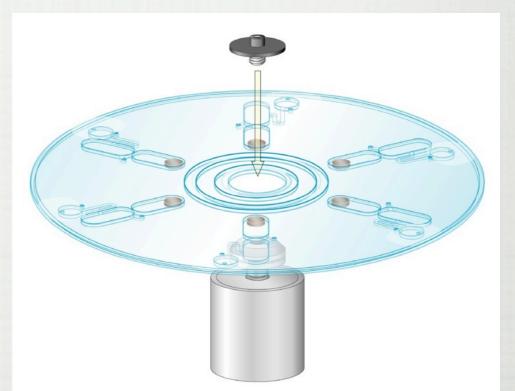






# CELL LYSIS ON CD

- LYSIS IS IMPORTANT STEP IN SAMPLE PREPARATION, NEED TO GET GENETIC MATERIAL OUT OF CELL/SPORE FOR DOWNSTREAM NUCLEIC ACID ANALYSIS STEPS
- MANY DIFFERENT KINDS OF LYSIS ARE USED: MECHANICAL, CHEMICAL, THERMAL, ETC
  - KIDO, ET AL UTILIZED MECHANICAL BEAD-BEATING LYSIS BY USING MAGNETIC DISCS WITHIN A CD DEVICE TO CREATE SHEER FORCES THAT LYSE THE CELLS
    - RAPID-- LYSIS OCCURS WITHIN 3-5 MIN
    - DOWNSTREAM PROCESSING STEPS



H. KIDO





# CELL LYSIS ON CD

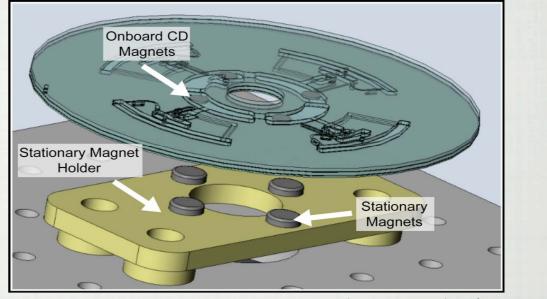
STEP	RPM	Seconds
Lysis	100	120
Transfer to clarification chamber	6000	30
Priming of siphon	100	10
Transfer to collection chamber	1500	10



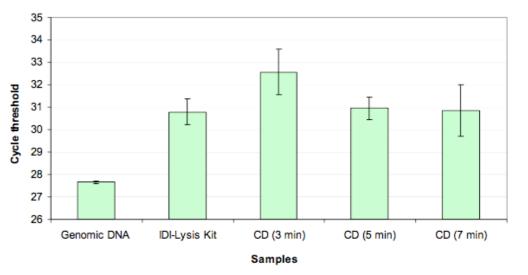


### CELL LYSIS ON CD

- IN THIS SYSTEM, A STATIONARY PLATFORM EMBEDDED WITH PERMANENT MAGNETS WAS ADDED BELOW THE SPINNING CD
- AS THE DISC SPINS, ON-CD MAGNETIC DISCS WERE ACTUATED, CAUSING GREATER IMPACTION THAN SUSPENDED BEADS ALONE. THE SYSTEM ALSO PERFORMED CLARIFICATION, WHEREBY LYSIS BY-PRODUCTS WERE CENTRIFUGED IN A CHAMBER, AND THE SUPERNATANT (CONTAINING THE RELEASED DNA) IS THEN SIPHONED INTO A SEPARATE COLLECTION CHAMBER.
  - LYSIS BOF B. GLOBIGII SPORES OCCURS FULLY AND RAPIDLY WHEN COMPARED TO PURIFIED GENOMIC DNA AND IDI LYSIS KIT



SIEGRIST, ET AL 2009



KIDO, ET AL 2006



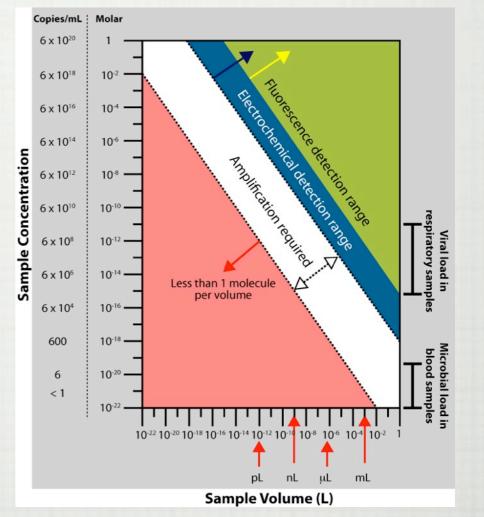


#### WORKING WITH LOW ANALYTE CONCENTRATIONS

ANALYTE CONCENTRATION DICTATES WHAT TYPE OF DETECTION METHOD TO USE

IN NA DIAGNOSTIC SYSTEMS, THE ANALYTE IS OFTEN PRESENT IN VERY LOW CONCENTRATIONS (<10 COPIES/ML) AS FOR THE CASE OF SEPSIS INFECTIONS

ARRAY HYBRIDIZATION IS A HIGHLY SENSITIVE AND SELECTIVE METHOD FOR VIRIAL OR MICROBIAL DETECTION FOR SAMPLES WITH LOW ANALYTE CONCENTRATIONS



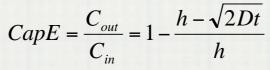
SIEGRIST, ET AL 2009





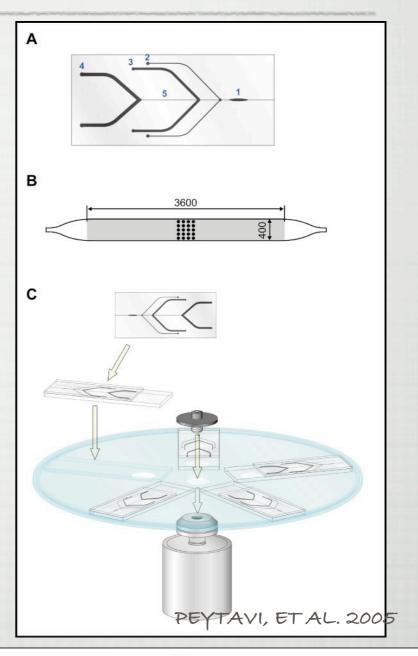
IN POC DIAGNOSTICS, A SPEEDY ANALYTICAL RESULT IS IMPORTANT, BUT PASSIVE (DIFFUSION-LIMITED) DNA HYBRIDIZATION MAY TAKE MANY HOURS

THE EQUATION DESCRIBING THE CAPTURE EFFICIENCY OF THE ARRAY IS:



WHERE COUT IS THE AVERAGE OUTPUT CONCENTRATION, CIN IS INPUT CONCENTRATION, H IS THE HEIGHT OF THE HYBRIDIZATION CHAMBER (50 MICRONS), T IS TIME AND IS CALCULATED FROM THE LINEAR VELOCITY AND THE SIZE OF THE ARRAY SPOT, AND D IS THE DIFFUSION COEFFICIENT OF DNA.

BASICALLY, WE WANT TO USE A MICROFLUIDIC FLOW CHAMBER TO SHORTEN THE DIFFUSION TIME OF THE TARGET MOLECULE TO ITS BINDING PARTNER



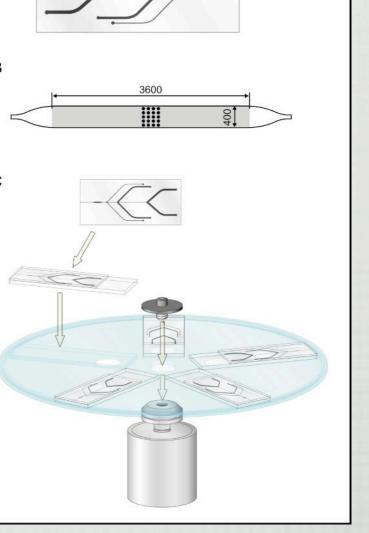




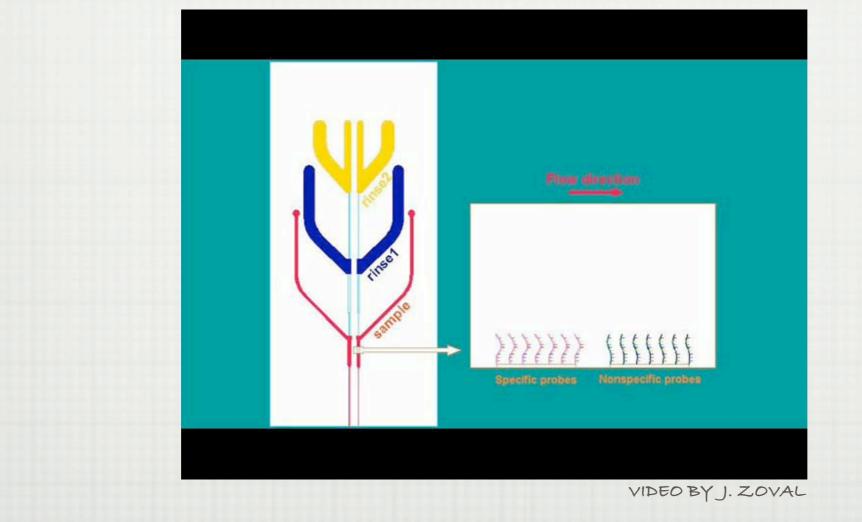
- Α WE DEVELOPED A SIMPLE MICROFLUIDIC SYSTEM MOLDED INTO A POLYDIMETHYLSILOXANE SUBSTRATE (PDMS) (PEYTAVI ET AL., 2005). PDMS IS A LOW-COST MATERIAL THAT CAN BE EASILY PROTOTYPED AND CAN MAKE REVERSIBLE WATERTIGHT SEALS WITH GLASS SLIDES. в THESE UNITS ARE PLACED ON AN ENGRAVED CD SUPPORT FIXED ON A ROTATIONAL DEVICE. IT COMPRISES A LINEAR HYBRIDIZATION CHAMBER COVERING THE MICROARRAY, 2 BUFFER RESERVOIRS, A SAMPLE RESERVOIR, A WASTE OUTLET, AND THE CONNECTING С CHANNELS. WASTE LIQUIDS ARE COLLECTED INTO A CIRCULAR CHAMBER MACHINED AROUND THE PLASTIC SUPPORT. CENTRIFUGAL FORCES DRIVE THE SAMPLE AND BUFFERS DIRECTLY ONTO THE MICROARRAY SURFACE. LOW DENSITY DNA PROBES MICROARRAY ARE PRINTED ON A STANDARD GLASS SLIDE MODIFIED WITH OUR NEW SURFACE CHEMISTRY, TO FORM A FUNCTIONAL MICROFLUIDIC UNIT.
  - UP TO 6 (150 SPOTS)-MICROARRAY SLIDES CAN BE HELD IN A SLIDE HOLDER CD

PEYTAVI, ET AL. 2005









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CAPILLARY VALVES BURSTAT: 450 RPM SAMPLE LOADING, 700 WASHING BUFFER, 850 RPM RINSE





Tuesday, March 16, 2010

Α

**B** 14000

12000

10000

9000 Black

4000

2000

PEYTAVI, ET AL. 2005

intensity

Fluorescence

S. aureus

14000

12000

10000

8000 -6000 -

4000-

0.

4500

4000

3500

3000 91 2500 -

1500

1000

500

saprophyticus probe

molyticus prob

Ŧ

aureus

S

S. epidermidis

epidermidis

S

10000

8000

6000

4000

2000

haemolyticus

S

S. haemolyticus

probe

emolyticus

Species-specific capture probes

#### DOWN TO 15 MINUTES

- THIS MICROFLUIDIC SYSTEM ALLOWS A DRASTIC REDUCTION IN THE VOLUME OF REAGENTS NEEDED FOR HYBRIDIZATIONS AND DOES NOT REQUIRE A PCR AMPLICON PURIFICATION STEP.
- OUR MICROFLUIDIC SYSTEM ALLOWED DETECTION OF PCR AMPLICONS GENERATED FROM 10 BACTERIAL GENOME COPIES, WHICH IS AT LEAST 1000 TIMES MORE SENSITIVE THAN RESULTS OBTAINED BY OTHER GROUPS USING MICROARRAY HYBRIDIZ,ATION ON MICROFLUIDIC DEVICES (LENIGK ET AL., 2002).
- IN TERMS OF SPECIFICITY, THIS SIMPLE SYSTEM DIFFERENTIATED 4 DIFFERENT STAPHYLOCOCCUS SPECIES WITH A POST-PCR HYBRIDIZATION PROTOCOL OF ONLY 15 MIN (PEYTAVI ET AL., 2005B





passive

saprophyticus

S

3500

3000

2500

1000

500

9 2000 1500 S. saprophyticus

nolyticus probe

epidermidis probe

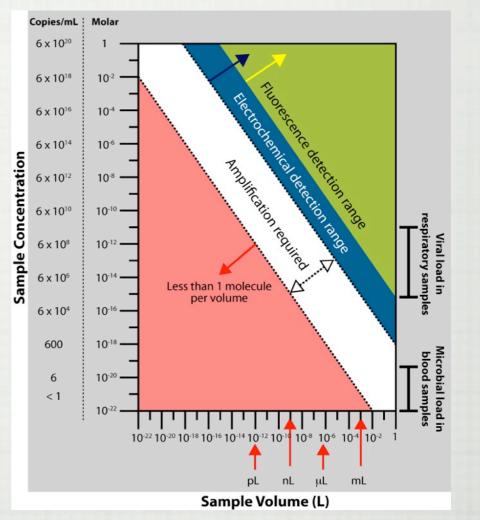
saprophyticus probe

□ flow-through

#### WORKING WITH LOW ANALYTE CONCENTRATIONS

- IN NA DIAGNOSTIC SYSTEMS, THE ANALYTE IS OFTEN PRESENT IN VERY LOW CONCENTRATIONS (<10 COPIES/ML) AS FOR THE CASE OF SEPSIS INFECTIONS
  - IF AN ASSAY REQUIRES 10 COPIES FOR NA DETECTION, THEN THE ASSAY REQUIRES 1 ML OF BLOOD SAMPLE

TO ACHIEVE DIAGNOSIS, THE DEVICE MUST BE ABLE TO PROCESS THESE LARGE SAMPLES, WHILE STILL ALLOWING FOR INTEGRATION OF DOWNSTREAM STEPS (CONCENTRATION, LYSIS, PCR, DETECTION)



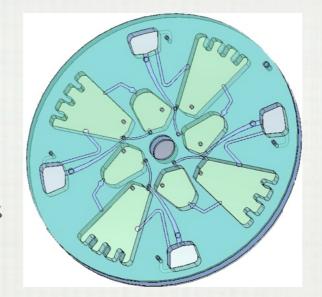
SIEGRIST, ET AL 2009

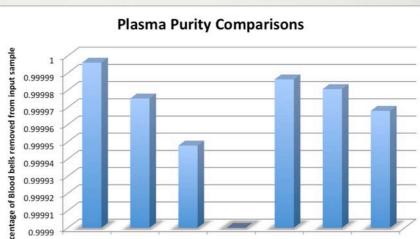




#### TOWARDS SAMPLE-TO-ANSWER SYSTEMS

WE HAVE DEVELOPED A LARGE-VOLUME BLOOD SAMPLE PREPARATION CD TO PROCESS THESE LARGER SAMPLE SIZES





2.5min

10min

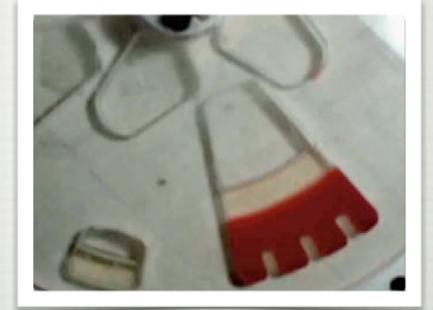
5min

2.5min

Tube 20min

PLASMA SEPARATION FROM 2 MLS OF WHOLE BLOOD

EQUIVALENT PURITY FROM CD SEPARATION @ 2.5 MIN COMPARED TO GOLD-STANDARD TUBE SEPARATION 10 MINUTES



M. AMASIA



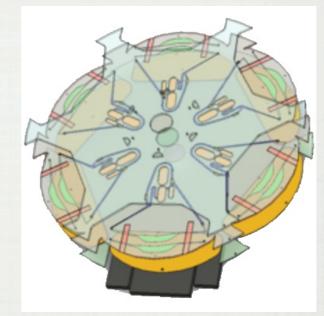


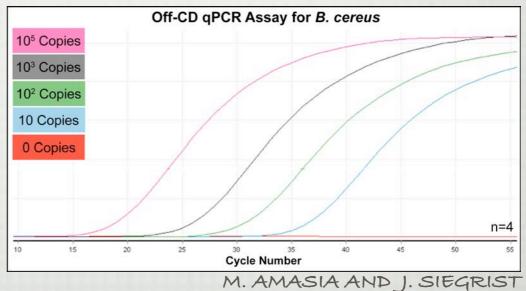
DEVELOPING SYSTEM FOR THE RAPID DETECTION OF B.ANTHRACIS BY REAL-TIME PCR.

SAMPLE PREPARATION (LYSIS AND CLARIFICATION, METERING)

PCR AMPLIFICATION

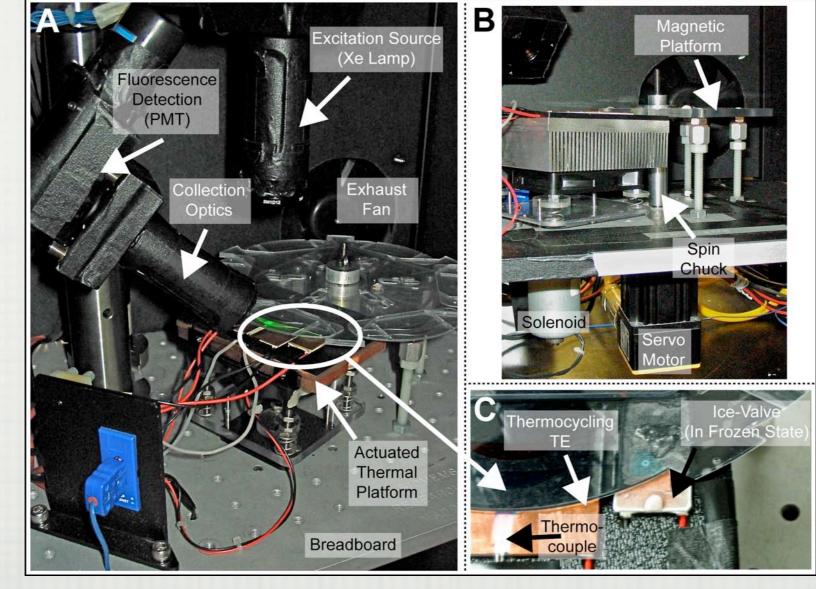
REAL-TIME PCR DETECTION











J.SIEGRISTAND M.AMASIA







Virus	NA Composition	Common Illness
Influenza A (various subtypes)	RNA	Common flu
Influenza B (various subtypes)	RNA	Common flu
H5N1 (Influenza A subtype)	RNA	Avian flu
H1N1 (Influenza A subtype)	RNA	Swine flu
SARS-CoV	RNA	SARS
HPIV (various subtypes)	RNA	Parainfluenza
Adenoviruses (various subtypes)	DNA	Various ailments

J. SIEGRIST



WORKING WITH M. BERGERON'S GROUP AT UNIVERSITY OF LAVAL TO DEVELOP A SYSTEM FOR RAPID RESPIRATORY VIRUS DETECTION THAT INTEGRATES:

SAMPLE PREPARATION (LYSIS AND CLARIFICATION, METERING)

D PCR AMPLIFICATION

DNA ARRAY DETECTION



