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Short Communication

Infection sustained by lineage B.1.1.7 of SARS-CoV-2 is characterised by longer persistence and higher viral RNA loads in nasopharyngeal swabs

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Highlights

- SARS-CoV-2 *lineage* B.1.1.7 is considered to have higher transmission capabilities, possibly related to higher viral loads in the upper respiratory tract or to a longer viral persistence of the infection in this tract.
- Statistically significant higher RNA loads were observed in nasopharyngeal swabs collected from individuals infected with *lineage* B.1.1.7 with respect to those infected by other SARS-CoV-2 lineages.
- A significant longer persistence of SARS-CoV-2 RNA was also observed in B.1.1.7-infected individuals

Abstract

Following the announcement on December 2020 about the emergence of a new variant (VOC 202012/01, B.1.1.7 lineage) in the United Kingdom a targeted surveillance was put in place in Abruzzo region (Italy), which allowed to detect 313 persons affected by lineage B.1.1.7, up to the 20th of February 2021. We investigated the results of RT-PCR on nasopharyngeal swabs tested from December 2020 to February 2021, to verify any difference on the viral load and persistence between people infected by lineage B.1.1.7 and others. Statistically significant lower values of C_T associated with the detection of the N protein encoding gene (C_{TN}) were observed in persons with lineage B.1.1.7 infection (median C_{TN} = 15.8) in comparison to those infected by other lineages (median C_{TN} = 16.9). A significant longer duration of the persistence of SARS-CoV-2 RNA in nasopharyngeal swabs was observed in persons with lineage B.1.1.7 infection (16 days) in comparison to those infected by other lineages (14 days).

Keywords: SARS-CoV-2; coronavirus; VOC 202012/01; lineage B.1.1.7; Abruzzo; Italy.

Introduction

Starting from March 2020, nasopharyngeal swabs collected in three provinces (Chieti, L'Aquila and Teramo) of Abruzzo, a central Region of Italy, are daily tested for the presence of SARS-CoV-2 RNA at the Istituto Zooprofilattico Sperimentale dell'Abruzzo e del Molise "G. Caporale" (IZSAM) (Danzetta et al, 2020).

Several SARS-CoV-2 variants are now circulating globally and some of them have raised international concern. One of these is certainly represented by the Variant of Concern (VOC) 202012/01 which belongs to the lineage B.1.1.7 (Rambaut et al., 2020). VOC 202012/01 is considered to have higher transmission capabilities (Davies et al, 2021), and although mechanisms underlying VOC 202012/01 spread are largely unknown, VOC 202012/01 seems to be associated with higher

viral loads and prolonged viral persistence in infected patients (Kissler et al., 2021). Furthermore, a six nucleotide deletion in S protein encoding gene of VOC 202012/01 is responsible for the S-gene drop out of a common used SARS-CoV-2 RNA real time-based detection kit (ThermoFisher, Waltham-MA, USA), which simultaneously detects two additional regions of SARS-CoV-2 genome namely ORF1ab and N protein encoding genes.

In order to unravel the spread of VOC 202012/01 in Abruzzo, a surveillance plan was established by the IZSAM. A two-step strategy was adopted (Bal et al., 2021). The first included a random selection of swabs resulting positive for SARS-CoV-2 RNA in a time period ranging from the beginning of December 2020 to February 20th 2021 but showing a readout pattern characterized by the S-gene drop-out. The second included whole genome sequence analysis of the S-negative swab samples with Threshold cycle (C_T) values ≤ 20 . Overall, at least the 10% of all positive samples were sequenced.

In the period of observation, 1,724 samples tested positive for SARS-CoV-2 RNA with the S-negative readout pattern. Of these, 655 were sequenced. VOC 202012/01 was detected in 313 individuals, mostly originating from the province of Chieti (n=258, 82.4%), which experienced an upsurge of COVID-19 cases in the first two months of the year 2021.

Hence, we deeper investigated molecular results of nasopharyngeal swabs tested from December 2020 to February 2021 to verify if VOC 202012/01 was associated to higher RNA loads and prolonged persistence in the respiratory tract with respect to those of other SARS-CoV-2 variants.

Methods

The workflow for SARS-CoV-2 RNA detection and sequencing have been previously described by our group (Danzetta et al. 2020). Sequences, once produced, are regularly shared with the GISAID database. For the estimation of the viral load in tested swabs, the C_T values associated with the detection of N protein encoding gene (C_{TN}) were chosen, since this protein is the less affected by the

mutation mainly targeting the S gene (Wu and Brian, 2010). The $C_T N$ gene values of the first positive nasopharyngeal swab of patients tested from December 2020 to February 2021 were analysed by comparing the median $C_T N$ values observed in the 313 VOC 202012/01-infected individuals with homologous values in individuals with S-positive results ($n= 2,344$). To obtain comparable sub-populations only those individuals with C_T values ≤ 20 were considered.

To verify any differences in the duration of the positivity at the molecular test, a further subset of the two before mentioned groups was defined. Reasonably, only those individuals with a final negative result, thus allowing to define the end of the positivity period, were included in the analysis. The difference (in days) between the date of first and the last positive nasopharyngeal swab was considered for each infected individuals.

The statistical analysis was performed using StatTools© (Palisade Corporation, Ithaca, NY, USA). A Mann–Whitney test was used to assess the statistical significance of differences among the $C_T N$ median values, whereas Chi-square and exact Fisher tests were used for comparing the percentages of people with clinical symptoms and died in the two groups. Level of statistical significance was set at 0.05.

Results and discussion

A statistically significant difference (Mann-Whitney Test, $P < 0.0001$) was observed between the median values of $C_T N$ observed in VOC 202012/01-infected individuals (median $C_T N = 15.8$) in comparison to S-positive infected individuals (median $C_T N = 16.9$) (Table 1). Furthermore, a statistically significant difference (Mann-Whitney Test, $P = 0.0317$) was observed between the median values of the duration of RNA positivity at the molecular test in VOC 202012/01-infected individuals ($n = 136$; median value = 16 days) in comparison to those of S-positive infected individuals ($n = 965$; median value = 14 days) (Table 1).

Viral load kinetics and the duration of viral shedding are important determinants for disease transmission (Cevik et al., 2020). In this regards also our analysis, performed in a given time period, suggests that VOC 202012/01 persists longer in the respiratory tract of infected individuals reaching higher RNA loads with respect to those of other SARS-CoV-2 variants. Although not a good predictor for viral load at individual level (Dahdouh et al., 2020), C_T values may give an indirect indication of the viral load in the population, as already seen in other studies (Veronesi et al., 2020; Hay et al., 2021).

One limitation of our study is the lack of information on the clinical status of all persons of two groups, which could be linked to different levels of C_T values and duration of the disease. However, the information about the presence or absence of clinical signs and the exitus of the disease was available for 140 VOC 202012/01-infected individuals and 961 S-positive infected individuals. In particular, 83.6% (C.I. 95%: 76.5%-88.8%) and 81.2% (C.I. 95%: 78.6%-83.5%) individuals showed COVID-19 clinical signs in VOC 202012/01-infected and S-positive infected persons, respectively. The difference about the two percentages is not statistically significant (Chi-square: 0.4686, $P = 0.4936$). Similarly, the difference between the percentages of deaths in the two groups, 2.1% (C.I. 95%: 0.8%-6.1%) for VOC 202012/01-infected and 4.1% (C.I. 95%: 3.0%-5.5%), was not significant (exact Fisher value = 0.3493). These findings, although limited to a sub-sample of the study population, suggest a similar clinical picture and gravity in the two populations.

Further analyses are reasonably warranted in order to establish the correlation between C_T values originating from infections with different variants and the presence of infectious (then transmissible) virus, the evolution of the spread of VOC 202012/01 in a given area and the impact on hospitalization and access to intensive care unit.

Conflict of interest

Authors declare no conflict of interests.

Conflict of interest

None.

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Ethical Approval

The results analysed in the present study derive from the official control activities performed by the Public Health Local Authority and no ethical approval is specifically requested.

References

Bal A, Destras G, Gaymard A, Stefic K, Marlet J, Eymieux S, Regue H, Semanas Q, d'Aubarede C, Billaud G, Laurent F, Gonzalez C, Mekki Y, Valette M, Bouscambert M, Gaudy-Graffin C, Lina B, Morfin F, Josset L; COVID-Diagnosis HCL Study Group. Two-step strategy for the identification of SARS-CoV-2 variant of concern 202012/01 and other variants with spike deletion H69-V70, France, August to December 2020. *Euro Surveill.* 2021 Jan;26(3):2100008. doi: 10.2807/1560-7917.ES.2021.26.3.2100008. PMID: 33478625; PMCID: PMC7848679.

Cevik M, Tate M, Lloyd O, Maraolo AE, Schafers J, Ho A. SARS-CoV-2, SARS-CoV, and MERS-CoV viral load dynamics, duration of viral shedding, and infectiousness: a systematic review and meta-analysis. *Lancet Microbe*. 2021 Jan;2(1):e13-e22. doi: 10.1016/S2666-5247(20)30172-5.

Epub 2020 Nov 19. PMID: 33521734; PMCID: PMC7837230.

Dahdouh E, Lázaro-Perona F, Romero-Gómez MP, Mingorance J, García-Rodríguez J. Ct values from SARS-CoV-2 diagnostic PCR assays should not be used as direct estimates of viral load. *J Infect*. 2020 Oct 24:S0163-4453(20)30675-7. doi: 10.1016/j.jinf.2020.10.017. Epub ahead of print. PMID: 33131699; PMCID: PMC7585367.

Danzetta ML, Amato L, Cito F, Di Giuseppe A, Morelli D, Savini G, Mercante MT, Lorusso A, Portanti O, Puglia I, Monaco F, Casaccia C, Di Gennaro A, Testa L, Migliorati G, D'Alterio N, Calistri P. SARS-CoV-2 RNA Persistence in Naso-Pharyngeal Swabs. *Microorganisms*. 2020 Jul 26;8(8):1124. doi: 10.3390/microorganisms8081124. PMID: 32722621; PMCID: PMC7466010.

Davies NG, Jarvis CI, Edmunds WJ, Jewell NP, Diaz-Ordaz K, Keogh RH. Increased hazard of death in community-tested cases of SARS-CoV-2 Variant of Concern 202012/01. *medRxiv* [Preprint]. 2021 Feb 3:2021.02.01.21250959. doi: 10.1101/2021.02.01.21250959. PMID: 33564794; PMCID: PMC7872389.

Hay JA, Kennedy-Shaffer L, Kanjilal S, Lennon NJ, Gabriel SB, Lipsitch M, Mina MJ. Estimating epidemiologic dynamics from cross-sectional viral load distributions. *medRxiv* [Preprint]. 2021 Feb 13:2020.10.08.20204222. doi: 10.1101/2020.10.08.20204222. PMID: 33594381; PMCID: PMC7885940.

Kissler S, Fauver JR, Mack C, Tai CG, Breban MI, Watkins AE, Samant RM, Anderson DJ, Ho DD, Grubaugh ND, Grad YH. Densely sampled viral trajectories suggest longer duration of acute infection with B.1.1.7 variant relative to non-B.1.1.7 SARS-CoV-2. Preprint, 2021.

<https://nrs.harvard.edu/URN-3:HUL.INSTREPOS:37366884>

Rambaut A, Loman N, Pybus O, Barclay W, Barrett J, Carabelli A, Connor T, Peacock T, Robertson DL, Volz E, on behalf of COVID-19 Genomics Consortium UK (CoG-UK). Preliminary genomic characterisation of an emergent SARS-CoV-2 lineage in the UK defined by a novel set of spike mutations. 2020 <https://virological.org/t/563>.

Veronesi L, Colucci ME, Pasquarella C, Caruso L, Mohieldin Mahgoub Ibrahim M, Zoni R, Pergreffi M, Arcuri C, Seidenari C, Viani I, Capobianco E, Mezzetta S, Affanni P. Virological surveillance of SARS-CoV-2 in an Italian northern area: comparison of Real Time RT PCR cycle threshold (Ct) values in three epidemic periods. *Acta Biomed.* 2020 Jul 20;91(9-S):19-21. doi: 10.23750/abm.v91i9-S.10138. PMID: 32701912.

Wu HY, Brian DA. Subgenomic messenger RNA amplification in coronaviruses. *Proceedings of the National Academy of Sciences of the United States of America*, 2010, 107(27), 12257–12262. <https://doi.org/10.1073/pnas.1000378107>

Table 1. C_T N gene values of the first positive nasopharyngeal swab and persistence of positivity in patients tested from December 2020 to February 2021.

	C_T N values		Duration of positivity in nasopharyngeal swabs (days)			
	Patients with		Patients with			
	lineage B.1.1.7 (n=313)	Others (n=2344)	lineage B.1.1.7 (n=136)	Others (n=965)		
Mean	15.4	16.4	17.4	17.1		
Median	15.8	16.9	P < 16.0	14.0	P <	
2.5 percentile	9.6	10.4	0.0001 7.0	10.0	0.0317	
97.5 percentile	19.6	19.9	39.0	30.6		