Diversity and distribution of ticks from domestic ruminants in Lebanon

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Keywords

Domestic ruminants, Hard ticks, Ixodidae, Lebanon.

Summary

Ticks (Acari: Ixodidae) are ectoparasites infesting livestock in every geographic area in the world and they are vectors of several viral, bacterial, and protozoan pathogens to animals and humans worldwide. A deep knowledge of the geographical distribution of these arthropods would have a key role in the control of tick-borne diseases. Few data are available about tick presence in domestic ruminants in Lebanon. The study aimed at providing an analysis of tick presence and distribution in Lebanon. Ticks were collected from cattle, sheep, and goats farms distributed in 6 Lebanese provinces between June and September 2014. A total of 272 adult hard ticks were randomly collected from domestic ruminants (cattle, sheep, and goats) located at 37 Lebanese farms, distributed among 30 villages. Ticks belonged to 4 Ixodidae genera: Rhipicephalus (72.4%), Haemaphysalis (11.4%), Dermacentor (8.1%), and Hyalomma (8.1%). They included the following species: Rhipicephalus annulatus (50.7%), Rhipicephalus turanicus (18.8%), Hyalomma anatolicum (8.1%), Haemaphylasis punctata (11.4%), Dermacentor marginatus (8.1%), Rhipicephalus sanguineus (2.5%), and Rhipicephalus bursa (0.4%). Rhipicephalus turanicus and H. anatolicum were found on cattle, sheep, and goats, R. annulatus on cattle and sheep, R. sanguineus, D. marginatus and Hea. punctata on sheep and goats, while R. bursa was collected only on sheep. Tick species involved in pathogen transmission were found and some of the identified species were recorded in Lebanon for the first time.

Varietà e distribuzione di zecche in ruminanti domestici in Libano

Parole chiave

Ixodidae, Libano, Ruminanti domestici, Zecche dure.

Riassunto

Le zecche (Acari: Ixodidae) sono ectoparassiti in grado di infestare gli animali domestici in tutte le aree geografiche del pianeta e di trasmettere numerosi virus, batteri e protozoi patogeni sia per gli animali sia per gli esseri umani. Una conoscenza approfondita della distribuzione geografica di questi artropodi potrebbe avere un ruolo chiave nel controllo delle malattie trasmesse da zecche. Pochi dati sono disponibili riguardo alla presenza di zecche nei ruminanti domestici in Libano. Il presente studio ha avuto come obbiettivo analizzare la presenza e la distribuzione delle zecche in Libano. Le zecche sono state prelevate da bovini, ovini e caprini presenti in aziende libanesi, localizzate in 6 province. Il campionamento è stato effettuato nel periodo compreso tra giugno e settembre del 2014. Un totale di 272 esemplari adulti di zecche dure è stato prelevato in maniera casuale da ruminanti domestici (bovini, ovini e caprini) appartenenti a 37 aziende libanesi, localizzate in 30 villaggi. Le zecche appartengono a 4 generi di Ixodidae: *Rhipicephalus* (72,4%), *Haemaphysalis* (11,4%), *Dermacentor* (8,1%) e *Hyalomma* (8.1%). Le specie di zecche identificate includono: *Rhipicephalus annulatus* (50,7%),

Rhipicephalus turanicus (18,8%), Hyalomma anatolicum (8,1%), Haemaphylasis punctata (11,4%), Dermacentor marginatus (8,1%), Rhipicephalus sanguineus (2,5%) e Rhipicephalus bursa (0,4%). Rhipicephalus turanicus e H. anatolicum sono state prelevate da bovini, ovini e caprini; R. annulatus è stata rivenuta su bovini e ovini, R. sanguineus, D. marginatus e Hea. punctata su ovini e caprini; mentre R. bursa è stata riscontrata solo su ovini. Il presente studio ha permesso di riscontrare la presenza di alcune specie di zecca implicate nella trasmissione dei patogeni in Libano. Per alcune specie, la presenza in Libano è stata documentata per la prima volta.

Introduction

Ticks are obligate blood-sucking ectoparasites of mammals, birds, and reptiles. They affect human and animal health.

The impact of ticks is related to their ability to transmit diseases to humans and animals (Rajput 2006). Hence, ticks are responsible for severe economic losses in livestock both through direct and indirect effects. Direct effects can occur in several ways, as blood sucking can cause reduction in live weight, can limit livestock production, and can induce anaemia among domestic animals. Tick bites reduce the quality of hides and may cause irritation and serious physical problems to ruminants. Indirect effects can be due to the injection by certain ticks (Dermacentor andersoni) of a toxin causing paralysis (Durrey et al. 2012). In addition, ticks are vectors of several pathogens (virus, bacteria, protozoa, and filarial nematodes), which can cause diseases in animals and humans worldwide (Dantas-Torres 2008).

Approximately 878 tick species are known, the most of which belong to the 2 main families, *Ixodidae* and *Argasidae* (Anderson *et al.* 2008).

Due to their impact on human and animal health, as well on livestock production, the knowledge about the regional distribution and abundance of tick species is important.

Lebanon is located at the crossroads of the Mediterranean Basin and the Arabian hinterland. The Country belongs to the Mediterranean climatic zone, with rain falling during the winter and pronounced drought during the Summer. Studies on ticks prior to the present one have been conducted in Lebanon. However, the available information on tick distribution in Lebanon is scanty. Seven tick species have been reported to be endemic in Lebanon: Rhipicephalus annulatus (found on bovine in North Lebanon), Hyalomma aegyptium (on tortoise in Saida, Jezzine, Aatanit, Furzol, and in Batroun), Hyalomma sculzei (Camels without information about region distribution). No data have been reported about host and distribution for the 4 tick species, namely: Rhipicephalus sanguineus, Hyalomma anatolicum, Hyalomma excavatum and Dermacentor marginatus

(ElRai 2011, Apanaskevich et al. 2006, Hoogstraal et al. 1985, Otranto et al. 2014)¹.

The study aims to overcome the lack of data regarding tick presence on domestic animals in Lebanon by providing a survey of tick species presence and abundance in farms of ruminants in this country. A deeper understanding of the abundance of tick species involved in pathogens transmission, as well as their geographical distribution, is pivotal in controlling tick-borne diseases (TBDs) in the future.

Materials and methods

Geographic area and sampling period

The study was carried out into the following

¹ http://www.icttd.nl, http://www.phsource.us.

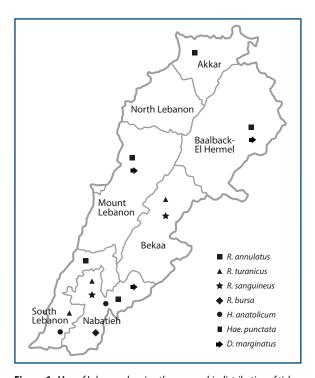


Figure 1. Map of Lebanon showing the geographic distribution of ticks in the 6 Lebanese provinces.

Nabatieh

South Lebanon

86.5%

82.4%

81.8%

3								
Provinces	Month	Average Minimum Temperature ± Standard deviation	Average Maximum Temperature ± Standard deviation	Average Minimum Relative Humidity	Average Maximum Relative Humidity			
Akkar June		16 ± 3.7	24.23±4.26	23%	93.5%			
Baalback-El Hermel	ck-El Hermel Sep 16.7± 1.45		30±2.34	46.2%	71.2%			
Mount Lebanon Sep		18.0±1.2	25.84±1.2	58.6%	94.1%			
Bekaa June 11.41+2.25		29.1+4.23	29.4%	66.3%				

 29 ± 3.6

29.35±1.44

28.6±2.85

Table 1. *Distribution of temperature and relative humidity in the 6 Lebanese provinces between June and September 2014.* Data obtained from the Lebanese Agricultural Research Institute.

6 Lebanese provinces: Akkar, Baalback-El Hermel, Bekaa, Mount Lebanon, Nabatieh, and South Lebanon (Figure 1). Being Lebanon characterised by a mild Mediterranean climate, tick collection was carried out in June 2014 in Akkar, Bekaa, Nabatieh and South Lebanon provinces, where late Spring and early Summer represent the period of tick activity. Moreover, in some farms of Nabatieh, a second tick sampling was also carried out in September 2014. In Baalback-El Hermel and Mount Lebanon provinces, where the weather is very moist, samples were collected in September 2014 (late Summer - early Autumn) (Table I).

 17.5 ± 1.5

18.5±1.3

21.15±1.17

Tick collection and storage

June

Sep

June

Ticks were collected from domestic animals (adult/young) in 37 farms distributed among 30 villages, where acaricide are not in use (Tables II and III). Farms were characterised by different kinds of breeding, including imported and local races, and mixed or mono species (Table IV).

Ticks were collected in the morning or in the evening on animals by using fine forceps avoiding crushing the arthropods (Walker *et al.* 2014). The arthropods were found mainly in the animal back, neck, ear, and mammary glands (Figures 2, 3, 4 and 5). Ticks collected from the host were stored in sealable vials containing 70% ethanol (Pérez-Eid 2009).

Tick genus and species identification

Adult hard ticks were identified under a stereomicroscope analysing the external morphological characteristic using morphological keys reported in literature (Hoogstraal *et al.* 1985, Walke *et al.* 2014, Pérez-Eid 2009).

Ticks were heated at 45°C in 10% KOH for a time ranging between 12 and 48 hours depending on the thickness of cuticle, which varies among the different tick species. Ticks were washed twice with distilled water and stained with Lugol's lodine for 15-30 minutes (Walker *et al.* 2014). Ticks were

then washed and fixed by using increasing grades of alcohol 20%, 40%, and 70% for 10 minutes, respectively (i.e. fixing the tick in each grade of alcohol for 10 minutes), and were subsequently mounted on glass slides in Hoyer's medium followed by hardening and drying at 45-50°C for 4-5 days. Morphologically identification of ticks was carried out using a microscope and reference taxonomic keys (Walker *et al.* 2014, Pérez-Eid 2009, Meddour *et al.* 2006, Hoogstraal *et al.* 1959, Kaiser *et al.* 1974).

21.1%

56.2%

25.6%

Table II. Distribution of farms in 6 Lebanese provinces.

Breeding	Provinces	Number of visited farms	Number of ticks in untreated farms	
	Akkar	4	25	
Bovine	Nabatieh	8	68	
Bovine	South Lebanon	2	17	
	Bekaa	0	0	
	Akkar	1	6	
	Baalback-El Hermel	1	7	
0 :	Bekaa	2	8	
Ovine	Mount Lebanon	1	3	
	Nabatieh	2	16	
	South Lebanon	0	0	
	Baalback-El Hermel	2	12	
Caprine	Mount Lebanon	3	22	
	Nabatieh	2	16	
Bovine/	Nabatieh	1	7	
Ovine	South Lebanon	1	10	
Bovine/	Akkar	1	6	
Caprine	South Lebanon	1	6	
Caprine/	Akkar	1	6	
Ovine	Nabatieh	2	25	
Bovine/ Caprine/ Ovine	South Lebanon	2	12	
Total		37	272	

In presence of ticks in a poor state of preservation or showing a doubtful morphological identification of species, a molecular identification of tick species was conducted. For this purpose, DNA was extracted from single specimens using the Pure link Genomic DNA Kit (Thermo Fisher™ Applied Biosystems™, Waltham, MA, USA) according to the manufacturer's instruction. A fragment of 360 base pairs (bp) of the mitochondrial small subunit 12SrRNA gene was amplified by polymerase chain reaction (PCR), the PCR primers were: forward primer T2A: 5'AAATGAGAGCGACGGGCGATGT3'and reverse primer T1B: 5'AAACTAGGATTAGATACCCT 3', followed by sequencing (Beati et al. 2001, Shemshad et al. 2011).

The PCRs were carried out in a final volume of $50 \mu l$, including: 50-100 ng of DNA, forward and reverse

Table III. Coordinates of the geographical Lebanese areas in which tick collection occurred.

Province	Place of collection	Longitude	Altitude	
	Adbel	34°32'6"N	36°57'50.4"E	300 m
	Bazbina	34°31' 0'' N	36°12'0"E	955 m
	El Kantara	34°31'33.078"N	36°00'3.0711"E	375 m
Akkar	Machha	34°32'25"N	36°7'56"E	349 m
	Machha	34°32'25"N	36°7'56"E	349 m
	Michmich	Michmich 34°21'24.0012"N		1,100 m
	Sahel Halba	34°33'2" N	36°4'41"E	120 m
	Chaat	35°00'30"N	36°00'36"E	1,000 m
Baalback- ElHermel	Chaat	35°00'30"N	36°00'36"E	1,000 m
Linemier	El Hermel	34°23′N	36°23′E	1,000 m
Bekaa	Zahle	33°50'48"N	35°54'07''E	963 m
DEKAA	Zahle	33°50'48"N	35°54'07''E	963 m
	Fakra	34°00'0.1002"N	35°48'18"E	1,140 m
Mount	Kfardibyan	34°00'39.90"N	35°49'29.38"E	1,200 m
Lebanon	Laklouk	34°7'59. 16"N 35°51'14.04"l		2,000 m
	WataHrajiliyi	34°00'1.032"N	35°47'36. 96"E	1,400 m
	AynEbel	33°00'42"N	35°14'24"E	800 m
	AynEbel	33°00'42"N	35°14'24"E	800 m
	AynEbel	33°00'42"N	35°14'24"E	800 m
	El Koulayaa	33°19'48"N	35°34'12"E	650 m
	El wazani	33°16'32"N	35°37'22"E	279 m
	El wazani	33°16'32"N	35°37'22"E	279 m
	El wazani	33°16'32"N	35°37'22"E	279 m
Nabatieh	Hasbaya	33°23'N	35°41′E	750 m
	Ibel El Saki	33°12'36"N	35°22'48"E	800 m
	Kafarkila	33°10'12"N 35°19'48"E		700 m
	Marjiiyoun	33°30'N	35°30'E	860 m
	Mayfadoun	33°20'9.6"N	35°27'43.2"E	470 m
	Rmeich	33°00'54"N	35°24"E	690 m
	Wata El Khiyam	33°19'37.8"N	35°36'40"E	700 m
	Zawtar El Charkieh	33°19' 33''N	35°28'34"E	475 m
	Ayn El Deleb	33°32'40.87''N	35°24'25.834''E	41 m
	Barich	33°16'22''N	35°21'9"E	358 m
South	El Bourghliyi	33°18'36''N	35°14'24"E	19 m
Lebanon	Maaroub	33°17'6"N	35°20'49.2"E	270 m
	Qinarit	33°30'17''N 35°22'44		233 m
	Zayta	33°30'29''N	35°23'03"E	300 m

primers 200 nM; each dNTPs 200 µM, 1.25 U of Taq Polymerase; MgCl₂ 1mM and the appropriate buffer 1X. Conventional PCR was carried out in a Thermo Fisher™ Applied Biosystems™ 2720 Thermal Cycler (Waltham, MA, USA). Thermal protocol included: an initial denaturation at 95°C for 15 minutes, followed by 35 cycles of denaturation

Table IV. Details of tick collection done from ruminants in 6 provinces of Lebanon.

Province	Collection Month in 2014	Kind of farms	Number of ticks	Tick species
		Bovine	25	R. annulatus
		Ovine	6	R. annulatus
Akkar	June	Ovine/Caprine	6	R. annulatus
		Bovine/ Caprine	6	R. annulatus
		Ovine	5	Hae. punctata
Baalback-	Cantambar		2	D. marginatus
El Hermel Caprine -	5	Hae. punctata		
		Сарппе	7	D. marginatus
Bekaa	luna	Ovina	ine 6 R. tu	
Dekaa	June	Ovine	2	R. sanguineus
		Ovine	3	Hae. punctata
Mount Lebanon	September	Canrina	9	Hae. punctata
LCDUIIOII		Caprine	13	D. marginatus
			57	R. annulatus
	June	Bovine	8	H. anatolicum
			3	R. turanicus
		Ovine	15	R. turanicus
			1	R. sanguineus
			7	R. turanicus
		Caprine	3	R. sanguineus
Nabatieh			2	H. anatolicum
	September	Caprine	4	Hae. punctata
		Bovine/Ovine	7	R. turanicus
			8	R. turanicus
	June	Caprine/Ovine	1	R. sanguineus
		Caprille/Oville	1	R. bursa
			10	H. anatolicum
	September	Caprine/Ovine	5	Hae. punctata
	June	Bovine	17	R. annulatus
		Bovine/Ovine/	7	R. annulatus
South		Caprine	5	R. turanicus
Lebanon		Paving /Ovin-	8	R. annulatus
		Bovine/Ovine	2	H. anatolicum
		Bovine/ Caprine	6	R. annulatus
Total			272	

at 95°C for 30 seconds, annealing at 51°C for 30 seconds, and elongation at 72°C for 1 minute. A final extension at 72°C for 5 minutes was carried out. The PCR products were visualized under UV light after electrophoresis on 1.5% agarose gel containing 1X SYBRSafe (Thermo Fisher™ Applied Biosystems™ Waltham, MA, USA). Amplicons were purified via vacuum filtration using the Machery Nagel NucleoFast 96 PCR Plate (Thomas Scientific, GmbH & Co. KG, Swedesboro, New Jersey, USA) following manufacturer's instructions. Sequencing reactions were carried out using the Big Dye Terminator Cycle Sequencing V1.1 (Thermo Fisher™ Applied Biosystems™ Waltham, MA, USA) in a 10 µl of final volume including BigDye™ Terminator v1.1/ v3.1 Sequencing Buffer (1X), primer 0.5 μM and about 10 ng of purified PCR products.

Sequencing reactions were purified using Sephadex plates (Millipore, Billerica, MA, USA) following



Figure 2. Ticks on mammary glands of cow.



Figure 4. *Ticks on the neck of cow.*

manufacturer's instructions and stored at +4°C until analysed. Sequencing reactions were performed on an ABI 3130X Genetic Analyser sequencer (Thermo Fisher™ Applied Biosystems™ Waltham, MA, USA), and sequence trace files were assembled using the multi-sequence alignment software Chromaspro (v.2.1.1). The sequences were compared with the ones present in Genbank using the Basic Local Alignment Search Tool.

Results

A total of 272 adult hard ticks were collected from domestic ruminants in 6 Lebanese provinces. Female



Figure 3. *Ticks in the back of cow.*



Figure 5. Two ticks in the ear of goat.

ticks were the 77.6% of the collected ticks, while male ticks represented the 22.4%. Ticks belonged to 4 genera, among which Rhipicephalus genus represented the highest frequent (72.4%) with respect to the other identified genera: Heamaphysalis, Dermacentor, and Hyalomma. Seven tick species were identified within the Ixodidae family (Table IV). Molecular analysis confirmed morphological identification for all the doubtful samples. Collected ticks included: Rhipicephalus turanicus and Hyalomma anatolicum, which were found on cattle, sheep, and goats; Rhipicephalus annulatus collected on cattle and sheep; Rhipicephalus sanguineus, Dermacentor marginatus, and Heamaphysalis punctata found on sheep and goats; while Rhipicephalus bursa was collected only on sheep (Table V).

One hundred and thirty-seven, 69, and 66 ticks were collected from cattle, sheep, and goats, respectively. The samples were spread across different Lebanese provinces as described below (Table VI). Rhipicephalus annulatus and H. anatolicum were the dominant ticks on cattle, R. turanicus and R. sanguineus were the prevalent species on sheep, while Hae. punctata and D. marginatus were the most frequently ticks collected on goats.

Out of the identified tick species, 5 where collected in June and – with the exception of *R. bursa* which were reported only at 650 m of altitude – the other 4 species occurred in both plain and mountainous areas. *Rhipicephalus annulatus* was found indeed in an altitude ranging from 19 m to 1,100 m, *R. turanicus* from 270 m to 963 m, *R. sanguineus* was collected in an altitude ranging from 270 m to 963 m, and *H. anatolicum* was found from 400 m to 860 m.

In September, other 2 tick species were collected in mountainous areas: *Hae. punctata* from 700 m to 2,000 m and *D. marginatus* from 1,000 m to 1,400 m.

Discussion

Lebanese provinces are characterised by a heterogeneous vegetation (pine, fir, oak, willow and cypress) and climate (temperature and humidity) and by the presence of livestock, which plays a very important role for the distribution, activity, growth, development, diversity, and reproduction of ticks.

The present study allowed for identifying 7 tick species in Lebanon (Table IV).

In particular, R. sanguineus is a monotropic (all developmental stages feed on the same host species), three-host (each life stage requires a new host to feed on) tick species (Dantas-Torres, 2010). It is typically located in warmer climates and its mainly host is dog, but it can feed also on ruminants. It has a hexagonal capituli and has been recorded worldwide, namely in India, Iran, China, Iraq, Bulgaria, and Europe (Kaiser et al. 1974, Lu et al. 2013, Ramezani et al. 2014, Shemshad et al. 2012, Toledo et al. 2009, Chhillar et al. 2014, Sarani et al. 2014, Yakhchali et al. 2011). Rhipicephalus annulatus is a typical one-host tick and its entire life cycle from larva to mated adult is confined to a single host and can be completed in 2 months (EFSA 2010) and its hosts are ruminants. Rhipicephalus annulatus can survive in mesomediterranean environment² and is distributed in Algeria, Iran, Senegal, and Ethiopia (Yakhchali et al. 2011, Mediannikov et al. 2014, Pegram et al. 1981, Benchikh-Elfegoun et al. 2007).

Table V. Distribution of hard ticks in different hosts in Lebanon.

Host	R. annulatus %	R. turanicus %	R. sanguineus %	R. bursa %	H. anatolicum %	Hae. punctata %	D. marginatus %
Bovine	44.45	2.22	0	0	3.68	0	0
Ovine	6.25	11.42	1.43	0	1.84	3.70	0.74
Caprine	0	5.16	1.07	0.40	2.58	7.70	7.36

Table VI. *Percentage of hard ticks scattered in different Lebanese provinces.*

Province	R. annulatus %	R. turanicus %	H. anatolicum %	Hae. punctata %	R. bursa %	R. sanguineus %	D. marginatus %
Nabatieh	23.50	14.70	7.40	3.30	0.40	1.80	0
South Lebanon	11.40	1.80	0.70	0	0	0	0
Akkar	15.80	0	0	0	0	0	0
Bekaa	0	2.30	0	0	0	0.70	0
Baalback-El Hermel	0	0	0	3.70	0	0	3.30
Mount Lebanon	0	0	0	4.40	0	0	4.80
Total %	50.70	18.80	8.10	11.40	0.40	2.50	8.10

Rhipicephalus bursa is a two-host species, 1 life cycle is completed in a year, it feeds on ruminants and horses² and it can survive in the Mediterranean climatic region. It is scattered in Algeria, Iran, Iraq, Bulgaria, and Europe (Shemshad et al. 2011, Shemshad et al. 2012, Toledo et al. 2009, Sarani et al. 2014, Benchikh-Elfegoun et al. 2007, Hasson et al. 2012, Arnaudov et al. 2014). Rhipicephalus turanicus is a three host-tick, ruminants and domestic animals are its hosts. It can survive in desert and Mediterranean climatic regions². It is distributed in Algeria, Iraq, and Europe (Shemshad et al. 2012, Pegram et al. 1981, Hasson et al. 2012). Haemaphysalis punctata is a three-host tick, ruminants, horses, and humans are its hosts and the entire life cycle can be completed in 1 year, although it usually takes 3 years². It is spread in Bulgaria, Algeria, Hungary, United Kingdom, and Ukraine (Yakhchali et al. 2011, Benchikh-Elfegoun et al. 2007, Arnaudov et al. 2014, Gyuranecz et al. 2011, Akimov et al. 2012).

Dermacentor marginatus is a three-host tick, the entire life cycle can be completed in 1 year and it can parasitize different kind of hosts, such as dogs, cattle, sheep, horses, and humans. This tick has 11 festoons, rectangular form of the basis captuli, short mouth, and large enamel on scutum. It inhabits pastures, temperate forest while it is common under oak and pine vegetation. Dermacentor marginatus is present in the Mediterranean region, having it been reported in Algeria, Bulgaria, and several European countries (Yakhchali et al. 2011, Benchikh-Elfegoun et al. 2007, Arnaudov et al. 2014, Tijsse-Klasen et al. 2015). Hyalomma anatolicum is a three-host tick, ruminants and camels are its hosts. It is widely distributed in central Asia, Middle and Near East, Arabia, South-Eastern Europe and North Africa (Apanaskevich et al. 2006).

Identified tick species are relevant since they can be vector of several pathogens. In particular, Rhipicephaline, as *R. annulatus* and *R. turanicus*, can transmit the etiological agents of babesiosis, theileriosis, anaplasmosis, and rickettsiosis. *Rhipicephalus bursa* can transmit babesiosis in small ruminants (*Babesia ovis*) and cattle (*Babesia bigemina*), anaplasmosis in bovine (*Anaplasma marginale*), and small ruminants (*Anaplasma ovis*) (EFSA 2010, Satta *et al.* 2011). *Rhipicephalus sanguineus* has been recorded worldwide and is able to transmit pathogens such as *Coxiella burnetii* and species belonging to *Rickettsia, Ehrlichia*, and *Anaplasma* genera (Dantas-Torres 2008, Sarih *et al.* 2005).

Haemaphysalis punctata can transmit agents of babesiosis and theileriosis both in cattle and sheep.

The tick was also associated with transmission of Tick borne encephalitis virus, Crimean-Congo haemorrhagic fever virus, and *Coxiella burnetii* (Satta *et al.* 2011).

Dermacentor marginatus can transmit several Babesia (Babesia canis, Babesia divergens, Babesia ovis, Babesia caballi), Theileria (Theileria ovis, Theileria equi) and Rickettsia (Rickettsia conorii and Rickettsia slovaca) species (Bonnet et al. 2013). This species has also been identified as one of the vectors of Coxiella burnetii (EFSA 2010).

Hyalomma anatolicum is considered vector of Crimean-Congo haemorrhagic fever virus, Theileria and Babesia species, Anaplasma marginale and arboviruses (Nabian et al. 2007).

This is the first survey of tick species infesting ruminants in Lebanon. A great diversity in tick species in domestic ruminants was reported and species that are known to transmit pathogens to animals and humans were found. Moreover, for the first time *Rhipicephalus turanicus* (Pomerantzev, 1940), *Rhipicephalus bursa* (Canestrini and Fanzago, 1878), and *Heamaphysalis punctata* (Canestrini and Fanzago, 1878) were detected in Lebanon.

Conclusions

The study permitted to identifying 7 tick species in domestic animals in Lebanon and it might serve as the starting point for future epidemiological studies.

Because of their ability to transmit pathogens to animals and humans, further studies are needed to analyse pathogens prevalence in Lebanese ticks. Characterization of tick-borne pathogens in their arthropod hosts will be relevant for epidemiological studies of tick-borne pathogens in Lebanon. Control of vector species abundance and distribution in association with the analysis of pathogen prevalence is essential to evaluate risks associated with pathogen transmission and to adopt suitable prevention measures to improve the health of both animals and humans.

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² http://www.icttd.nl.

References

- Akimov L. & Nebogatkin I. 2012. Distribution of the tick Haemaphysalis punctata (Acari, Ixodidae) in Ukraine. Vestnikzoologii, **46** (4), 46-51.
- Anderson J.F. & Magnarelli L.A. 2008. Biology of ticks. *Infect Dis Clin North Am*, **22** (2), 195-215.
- Apanaskevich D.A. & Horak I.G. 2006. The genus *Hyalomma* Koch, 1844. i. reinstatement of *Hyalomma* (euhyalomma) *glabrum* Delpy, 1949 (Acari, Ixodidae) as a valid species with a redescription of the adults, the first description of its immature stages and notes on its biology. *Onderstepoort J Vet*, **73** (1), 1-12.
- Arnaudov D.Y., Arnaudov A.D., Kirin D.A. & Gospodinova S.G. 2014. Ixodidae ticks of small ruminants in the region of Parvomai, Southern Bulgaria. *Bulgarian J Agric Sci*, **20** (3), 590-594.
- Beati L. & Keirans J.E. 2001. Analysis of the systematic relationships among ticks of the genera *Rhipicephalus* and *Boophilus* (Acari: Ixodidae) based on mitochondrial 12S ribosomal DNA gene sequences and morphological characters. *J Parasitol*, **87**, 32-48.
- Benchikh-Elfegoun M.C., Benakhla A., Bentounsi B., Bouattour A. & Piarroux R. 2007. Identification et cinétique saisonnière des tiques parasites des bovins dans la région de Taher (Jijel) Algérie. *Ann Méd Vét*, **151**, 209-214.
- Bonnet S., de la Fuente J., Nicollet P., Liu X., Madani N., Blanchard B., Maingourd C., Alongi A., Torina A., Fernández de Mera I.G., Vicente J., George J.C., Vayssier-Taussat M. & Joncour G. 2013. Prevalence of tick-borne pathogens in adult *Dermacentor* spp. ticks from nine collection sites in France. *Vector Borne Zoonotic Dis*, 13 (4), 226-236.
- Chhillar S., Chhilar J. & Kaur H. 2014. Investigations on some hard ticks (Acari: Ixodidae) infesting domestic buffalo and cattle from Haryana, India. *J Entomol Zool*, **2**, 99-104.
- Dantas-Torres F. 2008. The brown dog tick, *Rhipicephalus* sanguineus (Latreille, 1806) (Acari: Ixodidae): from taxonomy to control. *Vet Parasitol*, **152**, 173-185.
- Durrey J.K.T. 2012. Syndrome Paralysie dû aux morsures de tiques chez les ruminants. Thèse Doctorat Vétérinaire, École Nationale Vétérinaire d'Alfort, 13-36.
- European Food Safety Authority (EFSA). 2010. Panel on Animal Health and Welfare (AHAW). Scientific opinion on geographic distribution of tick-borne infections and their vectors in Europe and the other Regions of the Mediterranean Basin. *EFSA Journal*, **8** (9), 1723-1981.
- El Rai O. 2011. Enquête préliminaire sur les tiques et les piroplasmoses bovines au Liban-Nord. Thesis, Ecole Nationale De Médecine Vétérinaire De Sidi Thabet, 30-48.
- Gyuranecz M., Dénes B., Hornok S., Kovács G., Jurkovich V., Varga T., Hajtós I., Szabó R., Magyar T., Vass N., Hofmann-Lehmann R., Erdélyi K., Bhide M. & Dán Á. 2012. Prevalence of *Coxiella burnetii* in Hungary: screening of dairy cows, sheep, commercial milk samples, and ticks. *Vector Borne Zoonotic Dis*, **12** (8), 650-653.

- Hasson R. 2012. Tick distribution and infestation among sheep and cattle in Baghdad's south suburb. *Kufa J Vet Med Sci*, 3 (1), 77-90.
- Hoogstraal H. & Kaiser M.N. 1959. Observations on Egyptian *Hyalomma* ticks (Ixodoidea, Ixodidae). Biological notes and differences in identity of *H. anatolicum* and its subspecies *anatolicum* Koch and *excavatum* Koch among Russian and other workers. Identity of *H. lusitanicum*. *Ann Entomol Soc Am*, **52** (3), 243-261.
- Hoogstraal H. & Tatchell R. J. 1985. Ticks parasitizing livestock. *In* Ticks and tick-borne disease control. A practical field manual. *Tick Control*, **1**, 1-73.
- Kaiser M.N., Hoogstraal H. & Watson E.G. 1974. Ticks (Ixodoidea) on migrating birds in Cyprus, fall 1967 and spring 1968, and epidemiological considerations. *Bull Entomol Res*, **64** (1), 97-110.
- Lu X., Lin X., Wang J., Qin X., Tian J.H., Guo W.P., Fan F.N., Shao R., Xu J. & Zhang Y.Z. 2013. Molecular survey of hard ticks in endemic areas of tick-borne diseases in China. *Ticks Tick Borne Dis*, **4** (4), 288-296.
- Meddour Bouderda K. & Meddour A. 2006. Clés d'identification des Ixodina (Acarina) d'alergie. *Sciences & Technologie*, **24**, 32-42.
- Mediannikov O., Fenollar F., Socolovschi C., Diatta G., Bassene H., Molez J.F., Sokhna C., Trape J.F. & Raoult D. 2010. *Coxiella burnetii* in humans and ticks in Rural Senegal. *PLoS Negl Trop Dis*, **4** (4), 654-661.
- Nabian S., Rahbari S., Shayan P. & Haddad Zadeh H.R. 2007. Current status of tick fauna in north of Iran. *Iran J Parasitol*, 2 (1), 12-17.
- Otranto D., Huchet J.B., Giannelli A., Callou C. & Dantas-Torres F. 2014. The enigma of the mummy from Ancient Egypt and the origin of *Rhipicephalus sanguineus*. *Parasit Vectors*, **7** (2), doi: 10.1186/1756-3305-7-2.
- Pegram R.G., Hoogstraal H. & Wassef H.Y. 1981. Ticks (Acari: Ixodoidea) of Ethiopia. I. Distribution, ecology and host relationships of species infesting livestock. *Bull Entomol Res*, **71**, 339-359.
- Pérez-Eid C. 2009. Les tiques: identification, biologie, importance médicale et vétérinaire. Lavoisier, Paris, 188-254.
- Rajput Z.I., Hu S., Chen W., Arijo A.G. & Xiao C. 2006. Importance of ticks and their chemical and immunological control in livestock. *J Zhejiang Univ Sci B*, **7** (11), 912-921.
- Ramezani H., Riabi A. & Atarodi A. 2014. Faunistic study of hard ticks (Ixodidae) of domestic ruminants in the Southern Khorasan-e-Razavi in comparing with other regions of the province in 2012 Iran. *J Vet Adv*, **4** (5), 508-515.
- Sarani M., Telmadarraiy Z., Moghaddam A.S., Azam K. & Sedaghat M.M. 2014. Distribution of ticks (Acari: Ixodidae) infesting domestic ruminants in mountainous areas of Golestan province, Iran. *Asian Pac J Trop Biomed*, **4** (1), 246-251.
- Sarih M., M'Ghirbi Y., Bouattour A., Gern L., Baranton G. &

- Postic D. 2005. Detection and identification of *Ehrlichia* spp. in ticks collected in Tunisia and Morocco. *J Clin Microbiol*, **43**, 1127-1132.
- Satta G., Chisu V., Cabras P., Fois F. & Masala G. 2011. Pathogens and symbionts in ticks: a survey on tick species distribution and presence of tick-transmitted microorganisms in Sardinia, Italy. J Med Microbiol, 60, 63-68.
- Shemshad K., Rafinejad J., Kamali K., Piazak N., Sedaghat M.M., Shemshad M., Biglarian A., Nourolahi F., Valad Beigi E. & Enayati A.A. 2012. Species diversity and geographic distribution of hard ticks (Acari: Ixodoidea:Ixodidae) infesting domestic ruminants, in Qazvin Province, Iran. *Parasitol Res*, **110**, 373-380.
- Shemshad M., Shemshad K., Sedaghat M.M., Shokri M., Barmaki A., Baniardalani M. & Rafinejad J. 2012. First survey of hard tick (Acari: Ixodidae) on cattle, sheep and goats in Boeen Zahra and Takistan countries, Iran. *Asian Pac J Trop Biomed*, **2** (6), 489-492.

- Tijsse-Klasen E., Hansford K.M., Jahfari S., Phipps P., Sprong H. & Medlock J.M. 2015. Spotted fever group rickettsiae in *Dermacentor reticulatus* and *Haemaphysalis punctata* ticks in the UK. *Parasit Vectors*, **6**, 212.
- Toledo A., Jado I., Olmeda A.S, Casado-Nistal M.A., Escudero R. & Anda P. 2009. Detection of *Coxiella burnetii* in ticks collected from Central Spain. *Vector Borne Zoonotic Dis*, **9** (5), 465-468.
- Walker A.R., Bouattour A., Camicas J.L., Estrada-Peña A., Horak I.G., Latif A.A., Pegram R.G. & Preston P.M. 2014. Ticks of domestic animals in Africa: a guide to identification of species. Edinburgh, Bioscience Reports, 74-217.
- Yakhchali M., Rostami A. & Esmailzadeh M. 2011. Diversity and seasonal distribution of ixodid ticks in the natural habitat of domestic ruminants in north and south of Iran. *Rev Méd Vét*, **162** (5), 229-235.