

Hygienic characteristics and microbiological hazard identification in horse and donkey raw milk

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Veterinaria Italiana 2016, 52 (1), 21-29. doi: 10.12834/VetIt.180.545.1

Accepted: 15.02.2015 | Available on line: 31.03.2016

Keywords

Hazards,
Health,
Horse and donkey milk,
Hygiene,
Pathogens,
Risk assessment.

Summary

Today the interest toward horse (*Equus caballus*) and donkey (*Equus asinus*) milk for human consumption is receiving a renewed attention because of its particular composition, hypoallergenicity, and nutraceutical properties. The realistic perspective of global use of this aliment in balanced diets, especially for infancy and geriatrics, poses the need for a more in depth knowledge on milk hygiene and on the health status of dairy animals, as a prerequisite of consumers' safety. The aim of this paper was to review the available literature on the health and hygiene parameters as well as on the potential microbiological hazards in horse and donkey milk and the risks related to their consumption. Both microbial contamination and somatic cell count are reasonably low in equine milk and also the presence of pathogens, like *Escherichia coli* O157, *Salmonella* spp., *Campylobacter* spp., *Yersinia enterocolitica*, *Brucella* spp., *Mycobacterium* spp., *Bacillus cereus*, *Cronobacter sakazakii*, *Streptococcus equi* subsp. *zooepidemicus*, *Rhodococcus equi*, *Streptococcus dysgalactiae* subsp. *equisimilis*, *Clostridium difficile* and *Burkholderia mallei* is low. However, in those regions of the world where the prevalence of *Brucella* spp. and *Rhodococcus equi* is high, the alimentary risks could increase. Similarly, in areas with higher incidence of immunocompromised people, the increased risks should be warned not only for pathogens but also for opportunistic microbiota.

Caratteristiche igienico-sanitarie ed identificazione dei rischi microbiologici nel latte di cavalla e di asina

Parole chiave

Igiene,
Latte equino e asinino,
Patogeni,
Pericoli,
Salute,
Valutazione del rischio.

Riassunto

In questi ultimi anni il latte di cavalla (*Equus caballus*) e il latte di asina (*Equus asinus*) hanno fatto registrare un rinnovato interesse sia per la loro particolare composizione sia per le proprietà ipoallergeniche e nutraceutiche. La reale prospettiva per una diffusione globale di questo alimento innovativo, specialmente nell'ambito dell'alimentazione per i lattanti e gli anziani, richiede una più approfondita conoscenza dei pre-requisiti di sicurezza. Lo scopo di questa review è stato quello di esaminare i dati disponibili in letteratura circa i parametri igienico-sanitari e i potenziali pericoli microbiologici che possono riscontrarsi nel latte di cavalla e di asina e dei rischi connessi al consumo. Dai dati disponibili risulta che la contaminazione microbica e il numero delle cellule somatiche del latte sono ragionevolmente bassi e così pure la presenza di microrganismi patogeni, in particolare *Escherichia coli* O157, *Salmonella* spp., *Campylobacter* spp., *Yersinia enterocolitica*, *Brucella* spp., *Mycobacterium* spp., *Bacillus cereus*, *Cronobacter sakazakii*, *Streptococcus equi* subsp. *zooepidemicus*, *Rhodococcus equi*, *Streptococcus dysgalactiae* subsp. *equisimilis*, *Clostridium difficile* e *Burkholderia mallei*. Tuttavia, il livello di rischio alimentare potrebbe essere elevato in quelle regioni dove la prevalenza di *Brucella* spp. e di *Rhodococcus equi* è elevata. Inoltre, nelle aree con più alta presenza di persone immunocompromesse, un maggior livello di rischio è riconducibile non solo agli agenti patogeni, ma anche a quelli opportunisti.

Introduction

During the past few years the European scientific community has increasingly devoted more attention to the analysis of horse (*Equus caballus*) and donkey (*Equus asinus*) milk, mainly owing to both palatability and attractive nutrient and functional components that make it suitable to be used as an alternative food in the diet of infants with Ig-E mediated cow's milk protein allergy (Businco et al. 2000, Monti et al. 2007). Moreover, donkey milk can be considered a valuable food also for elderly consumers, due to its immunonutrient components (Salimei & Fantuz 2012). Equine milk is rich in lactose (61.0-65.8 g/kg milk), ω -3 (8.7-12.0% total fatty acids), and ω -6 polyunsaturated fatty acids (7.0-13.1% total fatty acids) (Pavlik et al. 2004). The macro mineral (calcium, potassium, phosphorus, magnesium) content is reported to be higher than in human milk, but lower than in ruminant milk; while sodium as well as zinc and manganese contents, are reported to be similar to human milk (Fantuz et al. 2012, Fantuz et al. 2013).

Lysozyme content in horse and donkey milk results to be high (mean value 600-4,000 mg/l) if compared to bovine (traces), caprine (traces), and human milk (Salimei et al. 2004, Guo et al. 2007, Uniacke-Lowe et al. 2010). Lysozyme, together with other factors, including immunoglobulins, lactoferrin, and lactoperoxidase, may function in the infant's digestive tract by reducing the incidence of gastro-intestinal infections and promoting proper healthy growth (Monti et al. 2007). The high lysozyme content, as revealed by previous studies, may contribute to the low bacterial concentration of properly milked equine milk (Salimei & Fantuz 2012). Also lactoferrin (average content 0.60-0.82 g/kg milk) is reported to have antimicrobial activity, among its possible functions, consistent with its digestibility (Uniacke-Lowe et al. 2010, Tidona et al. 2011).

It is worth noting that worldwide equine populations account for about 112 million heads, including 9% of mules. Horse and donkey populations are differently distributed among continents, ranging from 25-39% reported in Africa, Asia, and Americas to 6.3% in Europe and 0.4% Oceania (FAOSTAT 2013¹).

Donkey world population is estimated to be of 44 million, of which 40 million (96.4%) in the developing countries. Donkeys represent about 73% of Africa's total equine population, while they account for 49.0%, 15.2%, 8.2%, 2.2% in Asia, Americas, Europe, and Oceania, respectively (FAOSTAT 2013). Equine milk and meat are

traditionally consumed in many areas of the World as a dietary source of valuable proteins (Blench 2000, Benítez 2011), although cultural and technological constraints may limit their use. In Europe, the dairy donkey enterprise is developing in relatively few farms. One of the largest herds of dairy donkeys is located in Central Italy, with about 700 heads in a single farm. It is well known that milk for human consumption needs to be characterised by a low microbial contamination and by a low somatic cell count. Milk may in fact represent a microbiological risk especially when consumed raw. On this regard, the recent literature highlights cases of infections in humans as related to the consumption of raw milk from ruminants and caused by *Escherichia coli* O157, *Campylobacter* spp., *Salmonella* spp., and other pathogens or potential pathogen microorganisms (Claeys et al. 2013).

The aim of this article is to review the available literature on hygienic and safety characteristics of equine milk. A descriptive analysis of the main and potential microbiological hazards and their risk is therefore reported.

Hygiene and health parameters of horse and donkey milk

The limited data available in literature on horse and donkey milk show fairly variable levels of total microbial count (TMC) (3.40-5.87 log CFU/ml) (Figure 1). According to Šarić and colleagues (Šarić et al. 2012), breed and other animal factors, as well as interactivity of lysozyme and lactoferrin, may affect the antimicrobial properties of equine raw milk but more in depth details are lacking at the moment.

Studies carried out in Sicily (Italy) reported values of TMC between 4.15 and 5.63 log CFU/ml. Moreover, *Enterobacteriaceae* (0.68-1.93 log CFU/ml) and pathogens, or potentially pathogenic microorganisms such as *Staphylococcus aureus*, *Staphylococcus intermedius*, *Staphylococcus epidermidis*, *Streptococcus dysgalactiae* and *Cronobacter sakazakii* (formerly

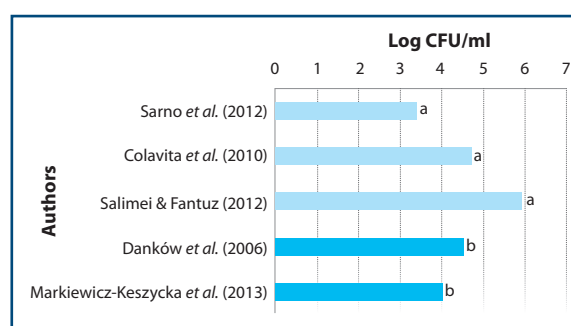


Figure 1. Total Microbial Count of donkey milk according to literature (mean values). a = Horse milk; b = Donkey milk.

¹ FAOSTAT, 2013. FAO Statistic Division. Available at <http://faostat.fao.org>

Enterobacter sakazakii), have been isolated in raw donkey milk (Conte et al. 2007).

During a 2 year study, 39 samples of bulk milk produced by 10 Martina Franca donkeys were evaluated for hygiene and health parameters. The value of the TMC at 30°C was 4.7 (\pm 0.73 s.d.) log CFU/ml and *Escherichia coli* was isolated in only 2 samples (2 log CFU/ml), while total coliforms showed counts ranging between less than 1.00 and 6.57 log CFU/ml. These data were strongly influenced by the values of 2 samples collected from a refrigerated tank, which had not been cleaned and sanitized properly, as verified later. This case underlined that, when hygiene is neglected, the level of microbial contamination can reach very high values in donkey milk. Pathogens, such as *Listeria monocytogenes* and *Salmonella* spp., have never been detected, yet; *Listeria* spp. were detected by polymerase chain reaction (PCR). This highlighted the importance of hygiene of both equipments and environment (Colavita et al. 2010a).

The wholesomeness of milk strictly depends on the mammary gland health. The values of Somatic Count Cells (SCC) ranged from 3.86 to 4.66 log/ml in one study (Conte et al. 2007), and 4.37 (\pm 0.32 s.d.) log/ml in another study (Colavita et al. 2010a) (Figure 2).

In a study on milk parameters over an entire lactation of jennies raised as companion animals, Pilla et colleagues (Pilla et al. 2010) noted a SCC content consistent with values observed in dairy jennies (Beghelli et al. 2009, Colavita et al. 2010 a, b). However, the number of SCC can be influenced by the milking intervals, as it remains almost unchanged for milking intervals of 3 and 5 hours but it increases at longer interval (D'Alessandro et al. 2009). It is important to note that horse and donkey need to be separated from the foal at least 3 hours before mechanical or manual milking. For this reason, data on donkey milk production are frequently expressed per 'milking session', i.e. from foal separation until donkeys leave the milking parlour (Salimei and Fantuz 2012). The low levels of SCC indicate an

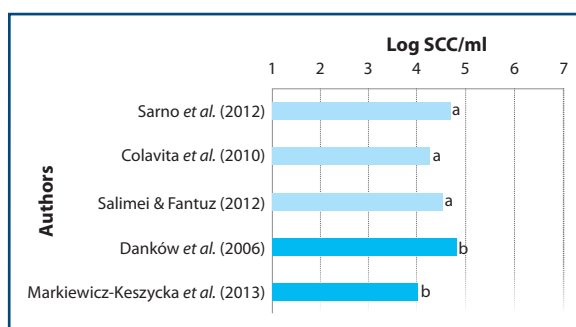


Figure 2. Somatic cell counts (SCC) of donkey milk (mean values) reported in literature. a = Horse milk; b = Donkey milk.

overall healthy status of mammary gland in the dairy equine where mastitis is less frequent than in the dairy cow (Svendsten 1997). This phenomenon is partly due to both the small volume of the mammary gland (max. 2-2.5 l) and its protected position so that teats are less exposed to traumas and infections. In addition, the mammary gland cistern has a limited capacity so that its emptying is guaranteed by foal suckling, when dams are not milked. Nonetheless, management of milking can affect the udder health. Clinical forms are often the result of traumatic lesions of teats caused by suckling or milking.

The prevalence of mastitis in donkeys is not known, with available data referring to mares: out of 100 lactating horses examined by Bartmann and colleagues (Bartmann et al. 1996), mastitis was diagnosed in 21 subjects. Although there are no data on the incidence of subclinical mastitis, these are considered more common than clinical mastitis. Perkins and Threlfall (Perkins and Threlfall 2002) reported *Streptococcus equi* subsp. *zooepidemicus* (32%), *Staphylococcus* spp. (15%), and *Actinobacillus* spp. (8.1%) as the main agents of mastitis in horses.

Cases of purulent mastitis caused primarily by streptococci and staphylococci have been reported in mares, glanders mastitis has rarely been reported, as well as mastites caused by *Mycobacterium avium* (Pavlik et al. 2004, Ryhner et al. 2009). In general, values of SCC/ml milk greater than 100,000 are considered pathological and index of mastitis. It is frequently possible to isolate *Streptococcus equi* subsp. *zooepidemicus* from mare udders, but pathogens may also be present in milk from healthy udders (Bohem et al. 2009). As Kocabiyik and colleagues (Kocabiyik et al. 2008) reported, out of 28 mares affected by mastitis, *S. equi* subsp. *zooepidemicus* was isolated in 37% of cases.

Analysis of microbiological hazards in horse and donkey milk

Since the data are currently scarce and fragmentary, it is not possible to approach properly the assessment of microbiological hazards associated with the consumption of equine milk. For this reason, in this context only a descriptive analysis of the actual and potential microbiological hazards and their risk is provided by considering both common risks with raw cow milk and specific ones for equids.

Escherichia coli O157 group and *Salmonella* spp.

With regard to microorganisms of the *E. coli* O157 group, the cow is considered the main reservoir, while

other species may serve as a secondary reservoir to humans, especially after contact with ruminants.

The few data available in literature show that the prevalence of *E. coli* O157 in horses is low. During a study conducted on 242 faecal samples of horses, the germ was isolated only in 1 subject (0.4%), which was farmed in promiscuity with cattle (Lengacher et al. 2010).

In some epidemiological cases, mixed farming can represent a risk factor, as evidenced by a survey in England and Wales, in which *E. coli* O157 was sought in the faeces of animals in some free stalls. The germ was found in 29% of the faeces samples of 365 cattle, in the 24% of the faeces samples from 426 sheep, in the 14.6% and 12.3% of the faeces samples of 26 donkeys and 85 horses, respectively (Pritchard et al. 2009).

In Germany, out of 400 samples of horse faeces, only in 1 case a strain of *E. coli* STEC belonging to serotype O113:H21 was identified by PCR, whereas all samples were negative for *Salmonella enterica* (Pichner et al. 2005).

Like other enteric pathogens, *Salmonella* spp. may contaminate the milk from faecal material during and after milking, although there are no reported cases of salmonellosis from consumption of equine raw milk so far (de Jager 2009). In studies carried out at slaughter on samples of faeces from horses coming from different European countries, *Salmonella* spp. was isolated in only 1.43% cases. However, this result may be misrepresented by the intermittent elimination of germs in the faeces (Colavita et al. 2010b).

Campylobacter spp. and Yersinia enterocolitica

Campylobacter spp. and *Yersinia enterocolitica* were found with low frequency in horse faeces (Browning et al. 1991). However, out of 100 samples of horse muscles from animals slaughtered in Piedmont (Italy), thermo-tolerant *Campylobacter* spp. were isolated in 3% cases. Outbreaks of campylobacteriosis caused by dairy products are most commonly associated with drinking unpasteurized milk. The infective dose of these bacteria is generally low. In European Union, the occurrence of *Campylobacter* ranges between 0% and 2.7% in raw milk and from 2.4% to 4.1% in cheese (EFSA 2012).

Moreover, *Yersinia enterocolitica* serotype O:5 was isolated in 9% of cases (Ercolini et al. 1997). Regarding the presence of these pathogens in milk, only data on cow's milk are available. In the same region of Italy on 30 samples of raw cow's milk, *Y. enterocolitica* was isolated in 4 cases (Franzin et al. 1984), while in Sicily it was detected in 8 samples of raw cow's milk out of 316 tested (Aiello et al. 1985). *Yersinia enterocolitica*

was found in 5% samples of bovine raw milk on 111 collected samples in Argentina (Mercado and Ibañez 1986). It is estimated that *Y. enterocolitica* is responsible for 2% of all infectious enteritis affecting children (Bottone 1999).

Brucellosis

In Europe, equine brucellosis is rare. Although there have been cases of infections by *Brucella suis* and *Brucella abortus* with rare episodes of abortion in the mare, and 2 cases of equine brucellosis caused by *B. suis* biovar 3 have been recently reported (Cvetnic et al. 2005).

The infection can cause mainly suppurative spinous bursitis and fistulizing. Brucellosis in horses has a prevalence of 5.8% in Egypt (donkey 7.3%), of 12.8% in India, and of 5.8% in Pakistan (Gul and Khan 2007). On a population of 346 horses and 28 donkeys, examined in Africa, 4.9% horses (17 subjects) and 3.6% donkeys (1 animal) were positive for brucellosis (Musa 2004). Moreover, on about 412 donkeys examined with seropositivity, a prevalence of 2.1%. *B. abortus* was reported (Elsalam Abdalla et al. 2010). Another sero-epidemiological survey carried out in Middle East reported that sero-prevalence for *Brucella* spp. was of 1% and 8.5% in horses and donkeys, respectively (Abo-Shehada 2009).

Mycobacteriosis

Tuberculosis in horses is extremely rare, but when it occurs it is often caused by *Mycobacterium avium* subsp. *avium* (agent of avian tuberculosis). Very sporadically both *Mycobacterium tuberculosis* and *Mycobacterium bovis* are the etiological cause of equine tuberculosis (Keck et al. 2010, Monreal et al. 2011). As also observed for *Rhodococcus equi*, the risk of transmission of tuberculosis infection from horses to humans can be considered very low, except for immunocompromised patients who are particularly susceptible to mycobacterial infection (Pavlik et al. 2004).

Bacillus cereus

Bacillus cereus causes 2 types of gastrointestinal disorders (diarrheic and emetic). The organism is ubiquitous in nature, and as a consequence, it is present in feedstuffs and contaminates raw milk from faeces, soil, and during processing. Its heat-stable spores survive to the pasteurization process. Several reports have shown that it can proliferate rapidly in milk, depending on storage temperature, and some authors indicated the release of enterotoxin in milk (Notermans et al. 2009). The contamination

of bovine milk by *B. cereus* is very common. Scatassa and colleagues (Scatassa *et al.* 2009) have isolated *B. cereus* at 3 log CFU/ml in samples of donkey milk. In the Netherlands, de Jager (de Jager 2009) analysed milk from 12 horse farms and found the organism in 2 samples, with counts ranging from 40 to 120 CFU/ml.

Cronobacter sakazakii

About 120-150 neonatal *Cronobacter* infections have been described as frequently associated with immunodeficiency syndrome in babies mainly fed with contaminated infant formula milk. Moreover, FAO/WHO cited cases of colonisation and infection with *Cronobacter* spp. in the elderly population. Microbiological criteria for food and process hygiene, relevant for the European Union, consider *Cronobacter* spp. as a possible risk factor in powdered infant formula and dried dietary foods for special medical purposes for infants of less than 6 months of age (Friedemann 2009). The pathogen was found in 2 samples of donkey milk (Conte 2007). However, the ingestion of low numbers of this micro-organism by healthy infants and children does not lead to illness (EFSA 2004).

Streptococcus equi subsp. zooepidemicus

Human infection by *Streptococcus equi* subsp. *zooepidemicus* is a known zoonosis, primarily in patients who consume milk or raw milk products (Barnham *et al.* 1989). There are numerous reported cases of illness, often severe (septicaemia, meningitis, abscess, fever), from the consumption of cow or goat raw milk. An episode caused by the consumption of unpasteurized cow's milk has been described in England, in which 11 people were affected and 7 died (Edwards *et al.* 1998). A case of streptococcal toxic shock syndrome (STSS) involving a woman suffering from meningitis caused by *S. equi* subsp. *zooepidemicus* and referable to the consumption of raw milk from mare has been reported in Japan (Hashikawa *et al.* 2004).

Rhodococcus equi

Rhodococcus equi causes pneumonia and enteritis with a high mortality rate in foals of 1-3 months. Isolated for the first time in 1923, it was named *Corynebacterium equi*. *Rhodococcus equi* is considered an important emerging pathogen in immunocompromised individuals, particularly if affected by Acquired Immune Deficiency Syndrome (AIDS), in which it plays the role of opportunistic

pathogen, with a high mortality rate. Eradication is difficult since it is transmitted by inhalation, through skin and tissues lesions as well as by food (Kedlaya *et al.* 2001). It lives and proliferates in soils rich in organic matter and it has been isolated from faeces of cattle, sheep, goats, pigs, and wild birds but, most frequently, from horse faeces (Takai *et al.* 1991). In foals it can be found at 3-4 log CFU/g, whereas in adults the counts are much lower. Thus, the foals are the true responsible for the maintenance of contamination at the farm level. In pigs and cattle, *R. equi* can cause injury, particularly at the level of the submaxillary lymph nodes, with some difficulty in the differential diagnosis with tuberculosis (Marcato 2008). In other species, albeit rarely, *R. equi* can give pyometra, mastitis, and purulent injuries (Cattabiani 2003); *R. equi* was also isolated from mammary quarters (13.7%) of dairy buffaloes (Bonini *et al.* 2007). The contamination of milk may be caused by environmental contamination or during suckling of the foal. While being a danger frequently present in some epidemiological conditions, the risk should be considered higher for immunocompromised consumers.

Streptococcus dysgalactiae subsp. equisimilis

Streptococcus dysgalactiae subsp. *equisimilis* comprises β -haemolytic streptococci belonging to the Lancefield group C. In humans, these colonize the pharynx, skin, gastrointestinal tract, and female genitourinary tract. This species causes severe pneumonia, septicemia, endocarditis, and toxic shock syndrome. In horses, *S. equisimilis* is generally considered infrequent or opportunistic, but it has recently been isolated from cases of strangles-like disease from aborted placentas and abscessed lymph nodes, suggesting a possible pathogenic role (Prezioso *et al.* 2010, Rantala *et al.* 2010).

Clostridium difficile

Clostridium difficile is a spore-forming Gram-positive, it produces toxins A and B, which cause gastroenteritis in humans with severe complications. It frequently develops resistance to antibiotics and causes outbreaks of nosocomial and community infections. It is assumed that aliments are a possible source of contamination, even though data are still inconsistent. *Clostridium difficile* has been isolated from foods commonly marketed in the United States, Canada, and Europe. The strains were often indistinguishable from those in humans, so that a common source or a zoonotic transmission is presumed. In 2005, *C. difficile* was isolated in 20% of beef samples in Canada and in 42% of beef and turkey

samples in the United States. In a survey carried out in Europe, *C. difficile* was isolated in 7.5% of samples of ready-to-eat foods. These data show that the germ is common in food and normal cooking of food will not idle the relatively rare spores. Although the issue is still controversial, some researchers believe that aliments of animal origin, including milk, may play a role in the transmission of *C. difficile* in humans, so that further studies are needed to design appropriate arrangements to control such transmission (Gould and Limbago 2010).

In 10% of horses *C. difficile* lives in the gut. Its presence may reach 45% in foals and in horses treated with antibiotics, but this germ rarely causes the disease, although in some cases colitis and diarrheal may arise. While the infection in humans is more common in hospitals, in equines it is found more frequently in the herd (Båverud *et al.* 2003).

Glanders

Burkholderia mallei is the causative agent of this disease that severely affects horses, mules, donkeys, and humans, with ulcerous lesions of the skin and lung that can even lead to death. Formerly widespread, the disease has been eradicated from many countries. Eliminated in Western Europe, glanders remains endemic in several countries in Asia, Africa, and South America (Wittig *et al.* 2006). In the last years, glanders outbreaks have been reported in Iraq, Iran, India, Pakistan, Turkey, and United Arab Emirates (Scholz *et al.* 2006). In Brazil, recent cases of glanders have been reported in some donkeys in the state of Pernambuco. The milk may be contaminated with material derived from pathological skin lesions (Mota *et al.* 2010).

Conclusions

Domestic equines are considered a dairy species worldwide since ancient times, but the recent scientific interest toward their possible role in human nutrition, mainly focused on nutritive, nutraceutical,

and hypoallergenic properties, is not yet accompanied by sufficient data on microbiological hygiene and health characteristics. If during milking the hygiene procedures are strictly observed, microbial contaminations of horse and donkey milk are generally low thanks to their anatomical and behavioural peculiarities. Both horses and donkeys exhibit high capacity of adaptation to severe environmental conditions: resistance to mammary gland infections is attested by the low somatic cell count of milk and rare presence of mastitis agents compared to cow milk.

According to the reviewed literature, hazard exposure associated to the consumption of raw horse and donkey milk has to be considered lower than for cow milk, especially for microorganisms like enterotoxigenic *E. coli* and thermo-tolerant *Campylobacter*. However, the hazards related to milk consumption vary depending on whether one considers risks for human or domestic animals health.

Even if the alimentary risks associated to equine milk consumption is reasonably low in Europe compared to bovine milk, thermal treatment before consumption is strongly recommended. The alimentary risks increase in those regions of the world where horse, donkey, and mules are still crucial components of micro-economies and prevalence of *Brucella* spp. and *R. equi* is higher.

For example, in those regions of the world where HIV/AIDS represent a serious health problem, the high incidence of the immunocompromised population warns on the increased risk for pathogens as well as for opportunistic microbiota possibly present in horse and donkey milk.

Acknowledgment

Part of the data reported in this paper have been published in Colavita G., Amadoro C & Salimei E. 2011. Sicurezza alimentare. Latte d'asina: aspetti igienico-sanitari e normativi. *Argomenti*, **13**, 61-70.

References

- Abo-Shehada M.N. 2009. Seroprevalence of *Brucella* species in equids in Jordan. *Vet Rec*, **165**, 267-268.
- Aiello P., Di Noto A.M. & Guercio M. 1985. Isolamento di *Yersinia enterocolitica* da latte crudo bovino. Proc. Soc Ital Buiatria, 589-591.
- Barnham M., Kerby J., Chandler R.S. & Millar M.R. 1989. Group C streptococci in human infection: a study of 308 isolates with clinical correlations. *Epidem Inf*, **102**, 379-390.
- Bartmann C.P., Blackmann E. & Klug E. 1996. Mammary diseases of the mare and possible consequences for the health of the foal. *Pferdeheilkunde*, **12**, 271-274.
- Båverud V., Gustafsson A., Franklin A., Aspán A. & Gunnarsson A. 2003. *Clostridium difficile*: prevalence in horses and environment, and antimicrobial susceptibility. *Equine Vet J*, **35**, 465-471.
- Beghelli D., Roscini A., Valiani A., Vincenzetti S., Cavallucci C. & Polidori P. 2009. Somatic (CSS) and differential cell count (DCC) during a lactation period in ass's milk. *Italian Journal of Animal Science*, **8**, 691-693.
- Benítez G. 2011. Animals used for medicinal and magico-religious purposes in western Granada Province, Andalusia (Spain). *J Ethnopharmacol*, **137**, 1113-1123.
- Blench R. 2000. A history of donkeys, wild asses and mules in Africa. In *The origins and development of Africa livestock: archaeology, genetics, linguistics and ethnography* (R. Blench & K.C. Mc Donald eds). Taylor & Francis, University College London Press, 339-354.
- Boehm K.H., Klug E. & Jacobs B.J. 2009. Mastitis in the mare - a long-term study on the incidence, clinical symptoms, diagnostics, microbiology, therapy and economic importance, as well as recommendations for veterinary practice. *Prakt Tierarzt*, **90**, 842-849.
- Bonini P.R., Mendoza-Sanchez G., Nader Filho A., Santos T.A.B., Langoni H., Tonhati H., Ferreira E.B.S., Ravena D.L., Sturion D.J. & Maia R.P. 2007. Distribution of contagious and environmental mastitis agents isolated from milk samples collected from clinically healthy buffalo cows between Brazilian dry and rainy seasons of the year. *Italian Journal of Animal Science*, **6**, 896-899.
- Bottone E.J. 1999. *Yersinia enterocolitica*: overview and epidemiologic correlates. *Microbes Infect*, **4**, 323-333.
- Browning G.F., Chalmers R.M., Snodgrass S.D., Batt R.M., Hart C.A., Ormarod S.E., Leadon D. & Thoneham S.J. 1991. The prevalence of enteric pathogens in diarrhoeic thoroughbred foals in Britain and Ireland. *Equine Vet J*, **23**, 405-409.
- Businco L., Giampietro P.G., Lucenti P., Lucaroni F., Pini C., Di Felice G. & Orlandi M. 2000. Allergenicity of mare's milk in children with cow's milk allergy. *J Allergy Clin Immunol*, **105**, 1031-1034.
- Cattabiani F. 2003. *Rhodococcus equi*: un patogeno opportunista di difficile eradicazione. *Annali Università Parma*, **23**, 233-241.
- Claeys W.L., Cardoen S., Daube G., De Block J., Dewettinck K., Dierick K., De Zutter L., Huyghebaert A., Imberechts H., Thiange P., Vandenplas Y. & Herman L. 2013. Raw or heated cow milk consumption: review of risks and benefits. *Food Control*, **31**, 251-262.
- Colavita G., Amadoro C., Maglieri C., Sorrentino E., Varisco G. & Salimei E. 2010a. Hygiene and health parameters of donkey's milk. EAAP, 61st Annual Meeting of the European Association of Animal Production, Crete, 44.
- Colavita G., Giaccone V., Miotti-Scapin R., Gemignani V., Dellamorte P., Chiarello M. & Mingarelli G. 2010b. Ricerca di *Salmonella* spp. nel contenuto intestinale di equini regolarmente macellati. Proc. LXIV Annual Meeting of the Italian Society for Veterinary Sciences (SISVET). Asti, Italy, 19-21 September 2010, 430-431.
- Conte F., Bargallo A. & Spanó G. 2007. Proposte per il controllo igienico-sanitario del latte d'asina in ambito di produzione primaria. Proc. XVII Congress of Italian Association of Veterinary Hygienists (AIVI). Cesenatico, Italy, 14-16 June 2007, 104-108.
- Cvetnic Z., Spicis S., Curic S., Jukic B., Loikic M., Albert D., Thiébaud M. & Gaurin-Bastji B. 2005. Isolation of *Brucella suis* biovar 3 from horses in Croatia. *Vet Rec*, **156**, 584-585.
- D'Alessandro A.G., De Petro R., Claps S., Pizzillo M. & Martemucci G. 2009. Yield and quality of milk and udder health in Martina Franca ass: effects of daily interval and time of machine milking. *Italian Journal of Animal Science*, **8**, 697-699.
- de Jager K.M. 2009. Safety of horse milk to humans and the effects of milking on the welfare of horses. Master thesis, University of Utrecht (<http://dspace.library.uu.nl/bitstream/handle/1874/33447/safety%20horse%20milk%2c%20laatste%20versie.pdf?sequence=2> accessed on 2 February 2016).
- Edwards A.T., Roulson M. & Ironside M.J. 1998. A milk-borne outbreak of serious infection due to *Streptococcus zooepidemicus* (Lancefield Group C). *Epidemiol Infect*, **101**, 43-51.
- Elsalam Abdalla M., Hassaballa Abdalla S. & Elzki R. 2010. Prevalence of *Brucella abortus* antibodies in donkeys in Gaderef State of Eastern Sudan. World Food System - A contribution from Europe. Tropentag, Zurich, 14-16 September 2010.
- Ercolini C., Serracca L., Migone L., Gorla M. & Ferrari A. 1997. Prevalenza di *Campylobacter* spp. *Yersinia enterocolitica*, *E. coli* O157:H7 in tessuto muscolare di bovino, suino, equino e cinghiale. *Il Progresso Veterinario*, **LXII**, 453-456.
- European Journal of Food Safety (EFSA). 2004. Microbiological risks in infant formulae and follow-on formulae. *EFSA J*, **113**, 1-35.
- European Journal of Food Safety (EFSA). 2012. The European Union summary report on trends and sources of zoonoses, zoonotic agents and food-borne outbreaks in 2010. *EFSA J*, **10** (3), 2597.
- Fantuz F., Ferraro S., Todini L., Piloni R., Mariani P. & Salimei E. 2012. Donkey milk concentration of calcium, phosphorus, potassium, sodium and magnesium. *Int Dairy J*, **24**, 143-145.
- Fantuz F., Ferraro S., Todini L., Piloni R., Mariani P. & Salimei E.

2013. Essential trace elements in milk and blood serum of lactating donkeys as affected by lactation stage and dietary supplementation with trace elements. *Animal*, **7**, 1893-1899.
- Franzin L., Fantino P. & Vidotto V. 1984. Isolation of *Yersinia enterocolitica*-like organisms from raw milk in Italy. *Curr Microbiol*, **10**, 357-360.
- Friedemann M. 2009. Epidemiology of invasive neonatal *Cronobacter* (*Enterobacter sakazakii*) infections. *Eur J Clin Microbiol*, **28**, 1297-1304.
- Gould L.H. & Limbago B. 2010. *Clostridium difficile* in food and domestic animals: a new foodborne pathogen? *Clin Infect Dis*, **51**, 577-582.
- Gul S.T. & Khan A. 2007. Epidemiology and epizootiology of Brucellosis: a review. *Pak Vet J*, **27**, 145-151.
- Guo H.Y., Pang K., Zhang X.Y., Zhao L., Chen S.W., Dong M.L. & Ren F.Z. 2007. Composition, physiochemical properties, nitrogen fraction distribution, and amino acid profile of donkey milk. *J Dairy Sci*, **90**, 1635-1643.
- Hashikawa S., Linuma, Y., Furushita M., Ohkura T., Nada T., Torii K., Hasegawa T. & Ohta M. 2004. Characterization of group C and G streptococcal strains that cause streptococcal toxic shock syndrome. *J Clin Microbiol*, **42**, 186-192.
- Keck N., Dutruel H., Smeyi F., Nodet M. & Boschiroli M.L. 2010. Tuberculosis due to *Mycobacterium bovis* in a Camargue horse. *Vet Rec*, **166**, 499-500.
- Kedlaya I., Ing M.B. & Wong S.S. 2001. *Rhodococcus equi* infections in immunocompetent hosts: case report and review. *Clin Inf Dis*, **32**, 39-46.
- Kocabiyik A.L., Buyukcangaz E., Akkoc A., Ozakin C. & Cangul I.T. 2008. Disseminated *Streptococcus equi* subsp *zooepidemicus* infection in a foal and associated mastitis in a mare. *Turk J Vet Anim Sci*, **32**, 487-490.
- Lengacher B.K., Harpster T.R., Williams L., Lejeune M. L. & Jeffrey T. 2010. Low prevalence of *Escherichia coli* O157:H7 in horses in Ohio, USA. *J Food Prot*, **73**, 2089-2092.
- Marcato P.S. 2008. Patologia sistematica veterinaria, 2nd Ed. Edagricole, Bologna, 536 pp.
- Mercado E.C. & Ibañez S.B. 1986. Isolation of *Yersinia enterocolitica* from raw cow milk in Argentina. *Int J Food Microbiol*, **3**, 237-242.
- Monreal L., Segura D., Segalés J., Prades M. & Garrido J.M. 2011. Diagnosis of *Mycobacterium bovis* infection in a mare. *Vet Rec*, **149**, 712-714.
- Monti G., Bertino E., Muratore M.C., Coscia A., Cresi F., Silvestro L., Fabris C., Fortunato D. & Giuffrida M.G. 2007. Efficacy of donkey's milk in treating highly problematic cow's milk allergic children: an *in vivo* and *in vitro* study. *Pediatr Allergy Immunol*, **18**, 258-264.
- Mota R.A., Da Fonseca Oliveira A.A., Pinheiro Junior J.W., Da Silva L.B.G., Brito M.F. & Rabelo S.S.A. 2010. Glanders in donkeys (*Equus asinus*) in the state of Pernambuco, Brazil: a case report. *Braz J Microbiol*, **49**, 146-149.
- Musa M.T. 2004. A serological study on equine brucellosis in Darfur, Western Sudan. *The Sudan J Vet Res*, **19**, 7-11.
- Notermans S., Dufrenne J., Teunis P., Beumer R., Te Giffel M. & Peeters Weem P. 2009. A risk assessment study of *Bacillus cereus* present in pasteurized milk. *Food Microbiol*, **14**, 143-151.
- Pavlik I., Jahn P., Dvorska L., Bartos M., Novotny L. & Halouzka R. 2004. Mycobacterial infections in horses: a review of the literature. *Vet Med Czech*, **49**, 427-440.
- Perkins N.R. & Threlfall W.R. 2002. Mastitis in mare. *Equine Vet Educ*, **5**, 99-102.
- Pichner R., Sander A., Steinruck H. & Gareis M. 2005. Occurrence of *Salmonella* spp. and shigatoxin-producing *Escherichia coli* (STEC) in horse faeces and horse meat products. *Berl Munch Tierarztl Wochenschr*, **118**, 321-325.
- Pilla R., Daprà V., Zecconi A. & Piccinini R. 2010. Hygienic and health characteristics of donkey milk during a follow-up study. *J Dairy Res*, **77**, 392-397.
- Prezioso S., Laus F., Tejada A.R., Valente C. & Cuteri V. 2010. Detection of *Streptococcus dysgalactiae* subsp. *equisimilis* in equine nasopharyngeal swabs by PCR. *J Vet Sci*, **11**, 67-72.
- Pritchard G.C., Smith R., Ellis-Iversen J., Cheasty T. & Willishaw G.A. 2009. Verocytotoxigenic *Escherichia coli* O157 in animals on public amenity premises in England and Wales, 1997 to 2007. *Vet Rec*, **164**, 545-549.
- Rantala S., Vähäkuopus S., Vuopio-Varkila J., Vuento R. & Syryänen J. 2010. *Streptococcus dysgalactiae* subsp. *equisimilis* bacteremia, Finland, 1995-2004. *Emerg Infect Dis*, **16**, 843-846.
- Ryhner T., Wittenbrink M., Nitzl D., Zeller S., Gygax D. & Wehrli Eser M. 2009. Infection with *Mycobacterium avium* subspecies *avium* in a 10 year old Freiberger mare. *Schweiz Arch Tierh*, **151**, 443-447.
- Salimei E. & Fantuz F. 2012. Equid milk for human consumption. *Int Dairy J*, **24**, 130-142.
- Salimei E., Fantuz F., Simoni A., Varisco G. & Chiari C. 2004. Ass's milking machine: first results on residual milk. Proc. 6th New Findings In Equine Practices Congress. Campobasso, Italy, 93-98.
- Šarić L.Ć., Šarić B.M., Mandić A.I., Torbica A.M., Tomić J.M., Cvetković D.D. & Okanović D.G. 2012. Antibacterial properties of domestic Balkan donkeys' milk. *Int Dairy J*, **25**, 142-146.
- Scatassa M.L., Arcuri F., Carrozzo A., Ducato B., Giosuè C., Lo Biundo G., Iannolino G., Arcuri L. & Mancuso I. 2009. *Bacillus cereus*: isolamento in latte di asina - nota preliminare. Proc. of LXIII Annual Meeting of the Italian Society for Veterinary Sciences (SISVET). Udine, Italy, 16-18 September, 410-412.
- Scholz H.C., Joseph M., Tomaso H., Al Dahouk S., Witte A., Kinne J., Hagen R., Wernery U. & Neubauer H. 2006. Detection of the reemerging agent *Burkholderia mallei* in a recent outbreak of glanders in the United Arab Emirates by a newly developed fliP-based polymerase chain reaction assay. *Diagn Microb Infect Dis*, **54**, 241-247.
- Svendsen E. 1997. The professional handbook of the donkey. In The donkey sanctuary, 2nd ed., Whitter Books Limited, London, 71-93
- Takai S., Ohbushi S., Koike K., Tsubaki S., Oishi H. & Kamada M. 1991. Prevalence of virulent *Rhodococcus*

- equi* in Isolates from soil and faeces of horses from horse-breeding farms with and without endemic infections. *J Clin Microbiol*, **12**, 2887-2889.
- Tidona F., Sekse C., Crescione A., Jacobsen M., Bordonarom S., Marletta D. & Vegarud G.E. 2011. Antimicrobial effect of donkeys' milk digested in vitro with human gastrointestinal enzymes. *Int Dairy J*, **21**, 158-165.
- Uniacke-Lowe T., Huppertz T. & Fox P.F. 2010. Equine milk proteins: chemistry, structure and nutritional significance. *Int Dairy J*, **20**, 609-629.
- Wittig M.B., Wohlsein P., Hagen R.M., Al Dahouk S., Tomaso H., Scholz H., Nikolaou K., Wernery R., Wernery U., Kinne J., Elschner M. & Neubauer H. C. 2006. Glanders-a comprehensive review. *Deut Tierarztl Woch*, **113**, 323-330.